

# Hidden layers of complexity

## How small RNA distribution and sample handling affect sequencing results

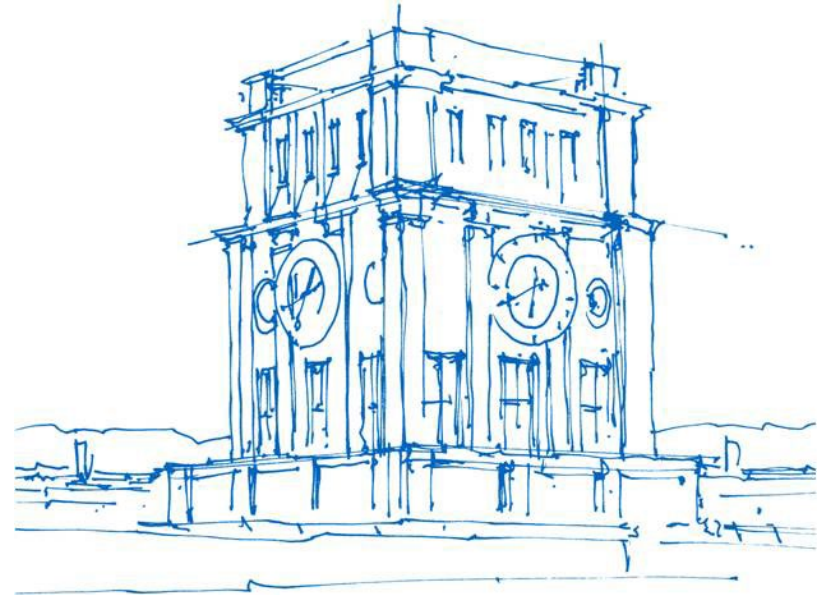
Dr. Benedikt Kirchner

Technische Universität München

School of Life Sciences

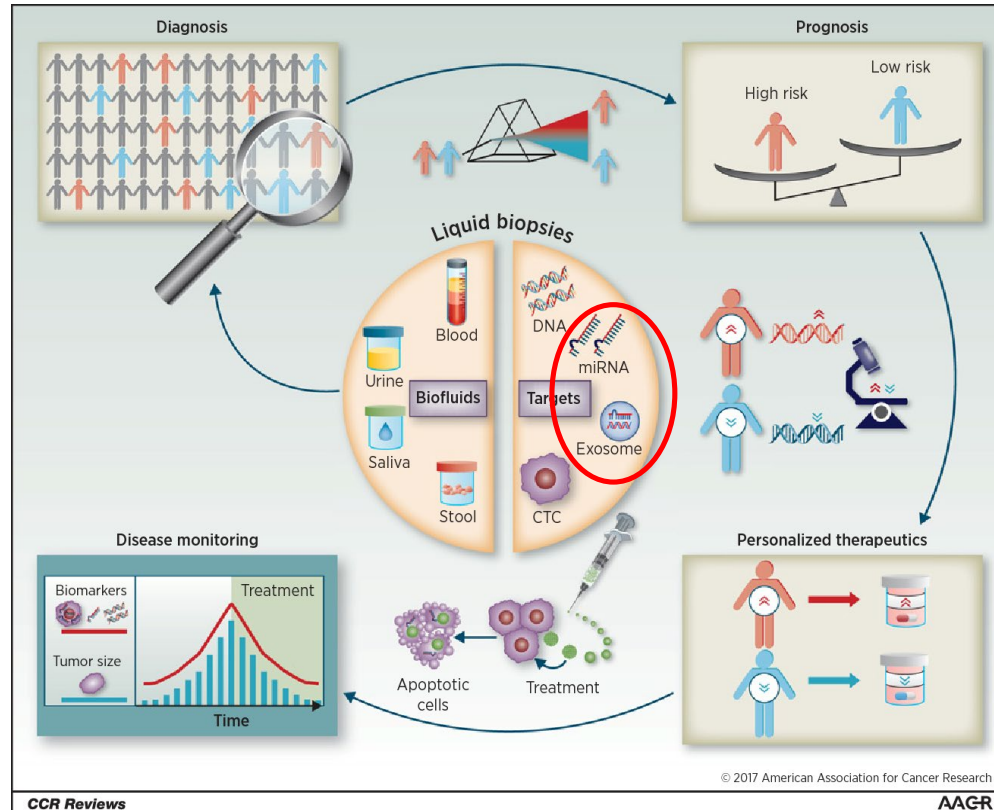
Division of Animal Physiology and Immunology

June 30<sup>th</sup>, 2023

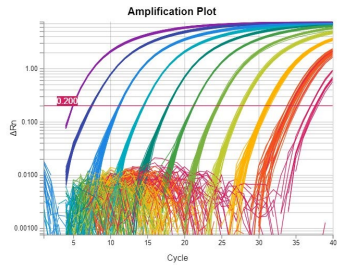
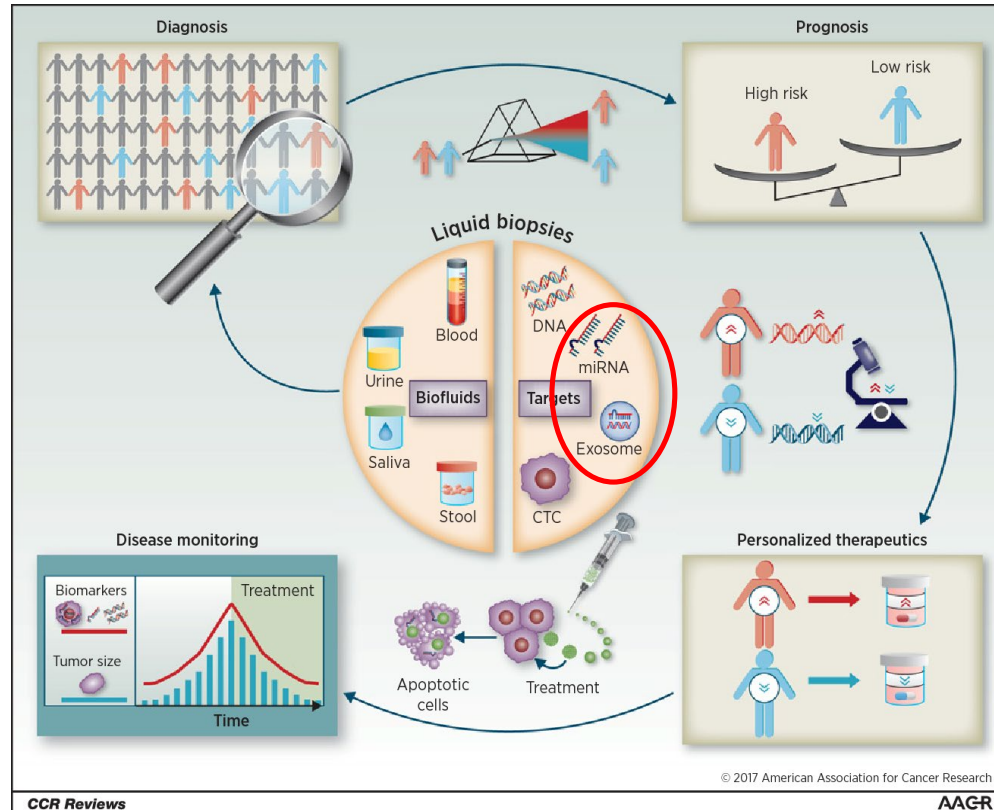


*Uhrenturm der TUM*

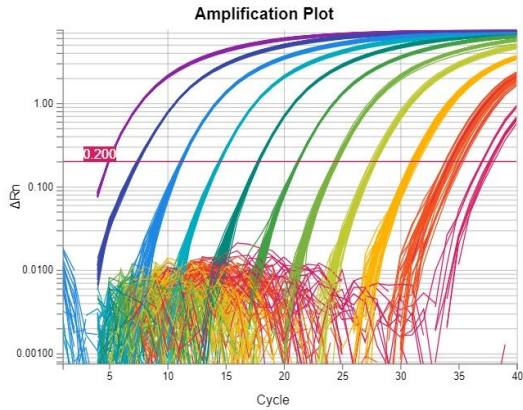
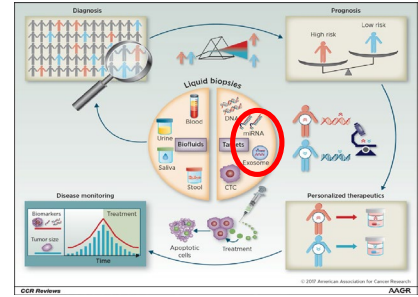
# Biomarker development in clinical settings



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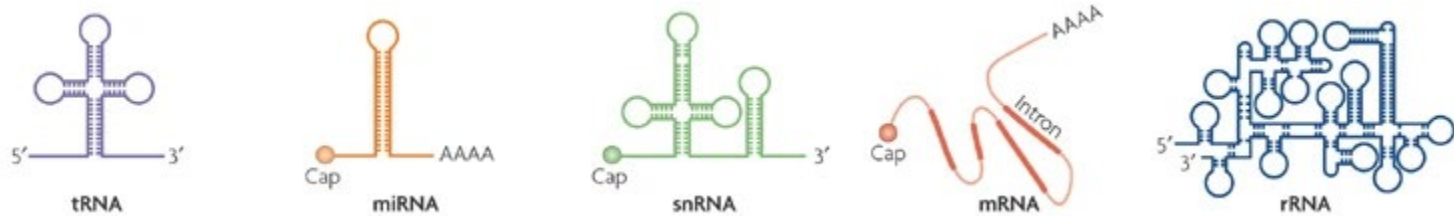
# Disagreement in methods



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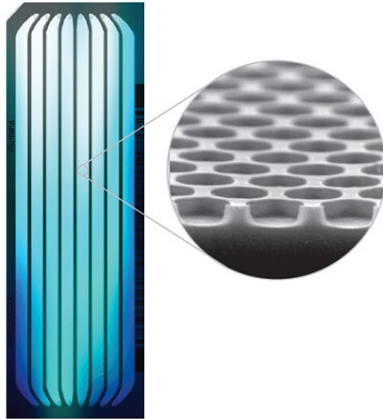
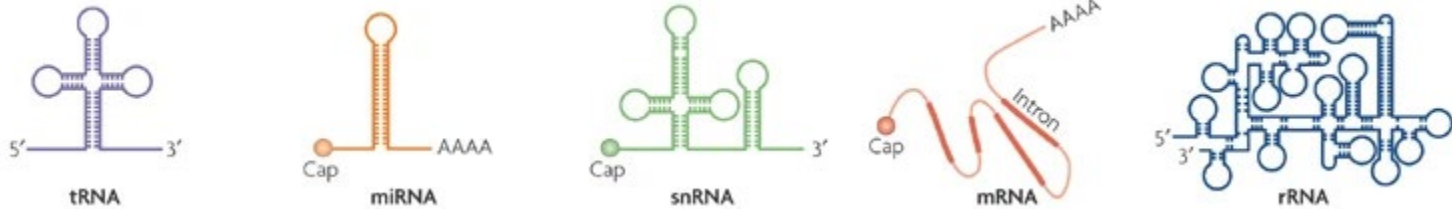


# Size selection bias in small RNA NGS



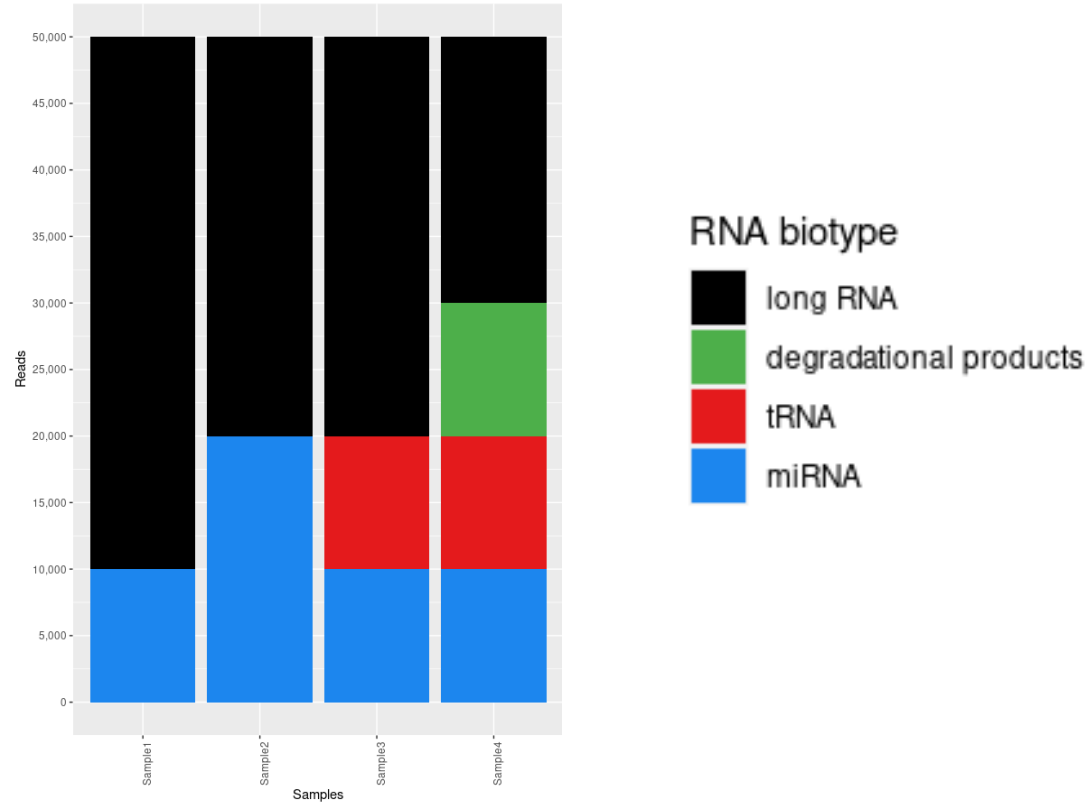
Enrichment of RNAs smaller than ~50 nt

# Size selection bias in small RNA NGS

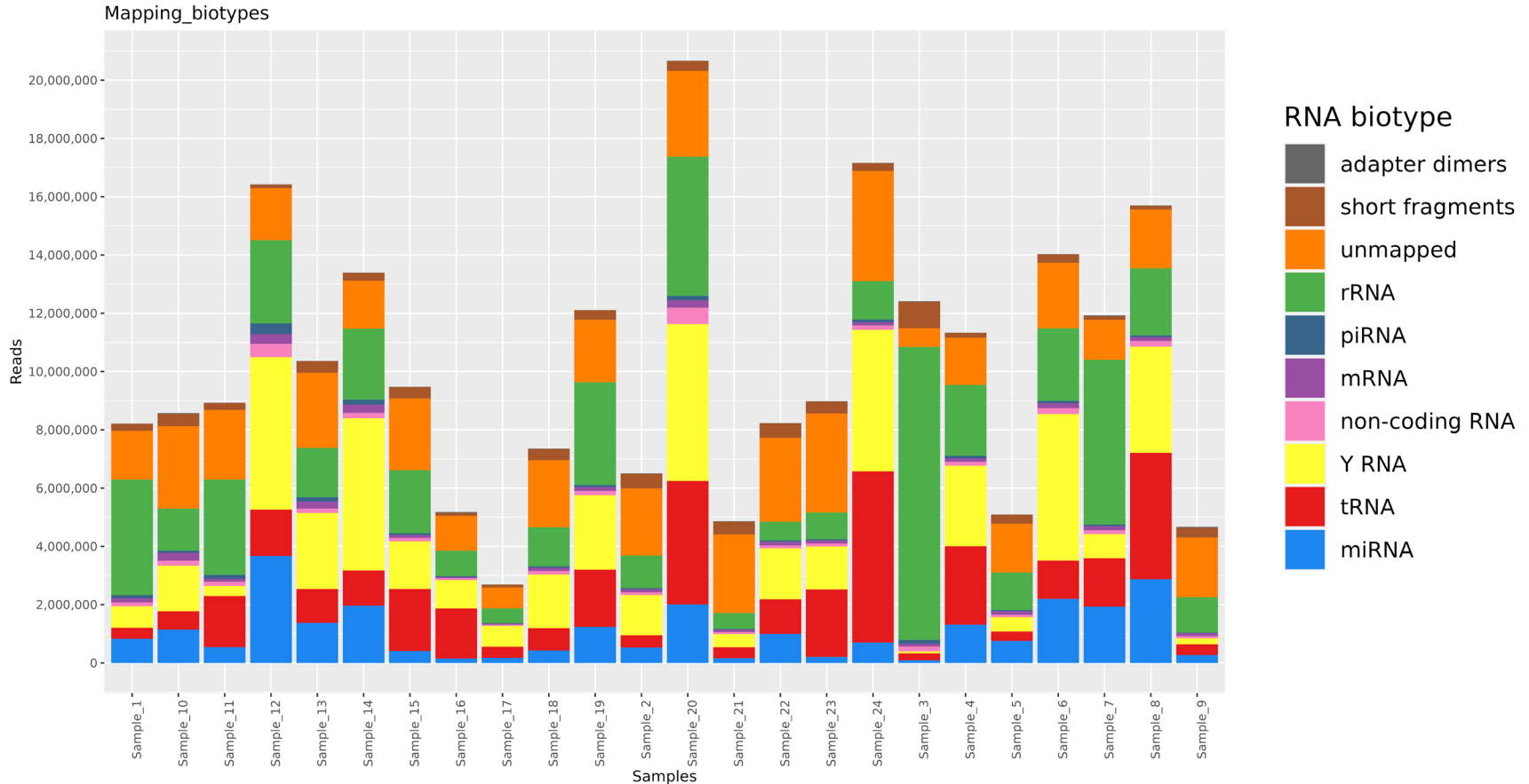


- Finite number of reads per flowcell
- Quantification always in relation to other small RNA transcripts
- Spike-in controls still rarely used

# Importance of normalization strategies

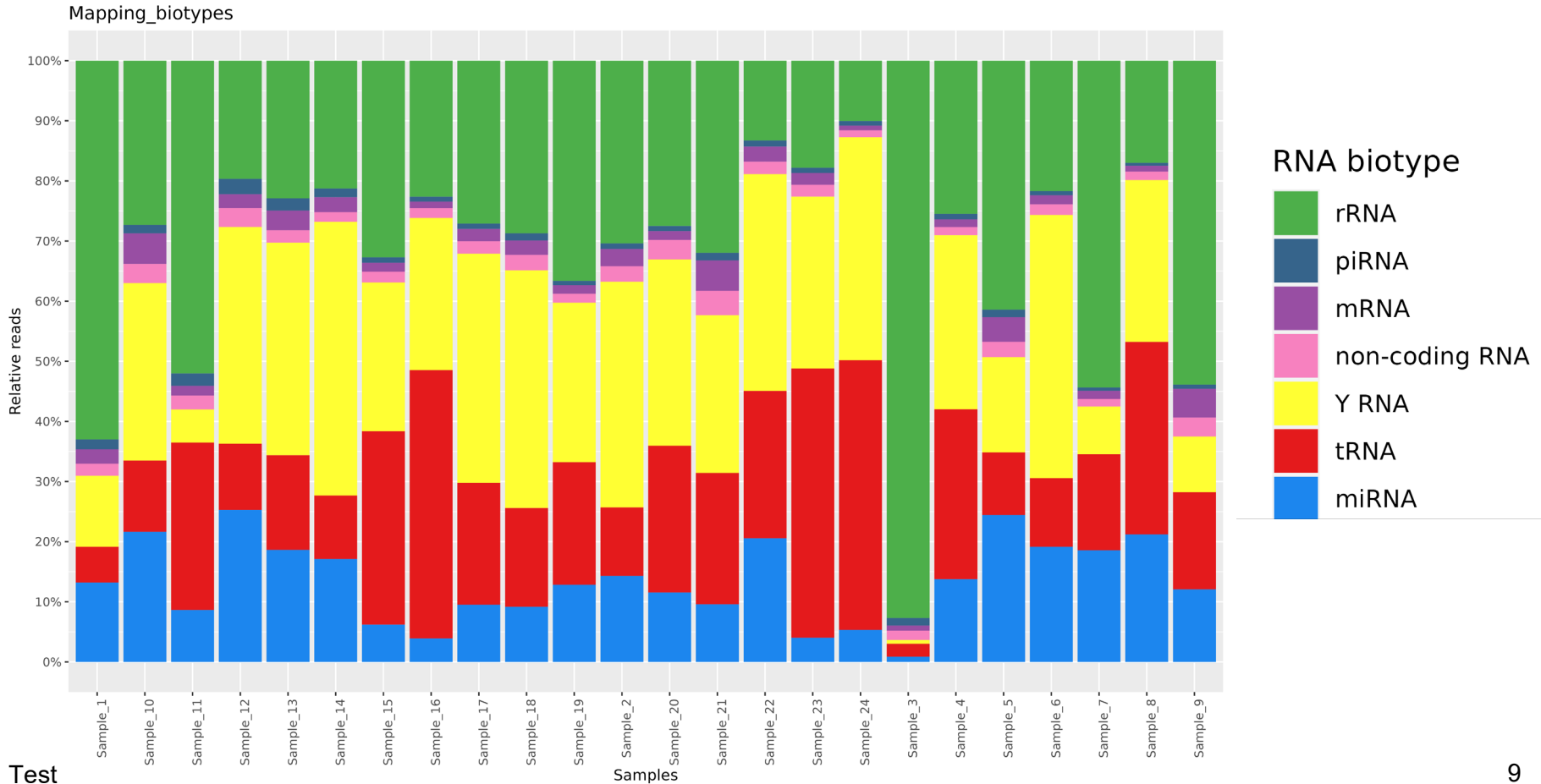


# Heterogenous biotype distribution

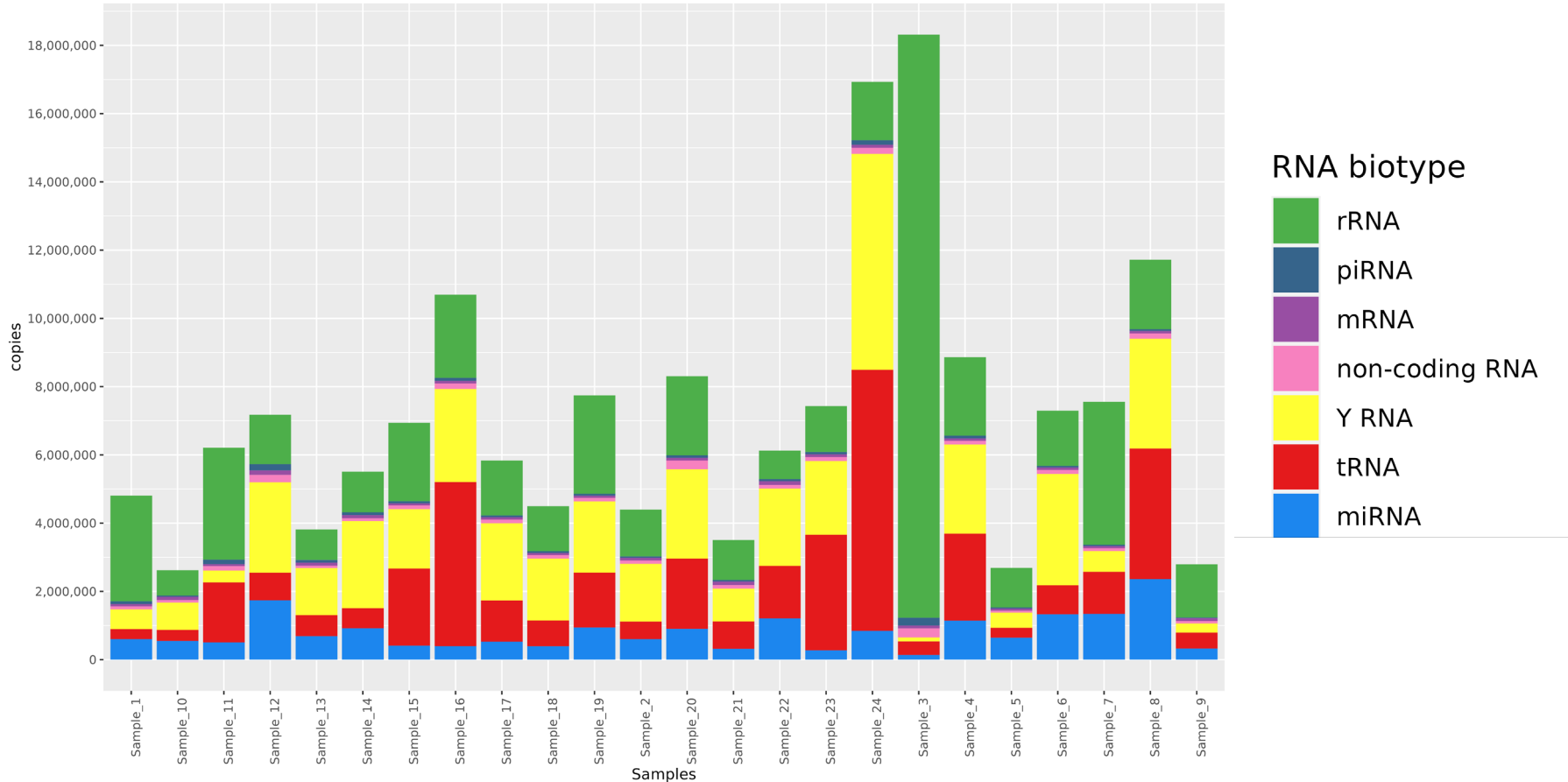




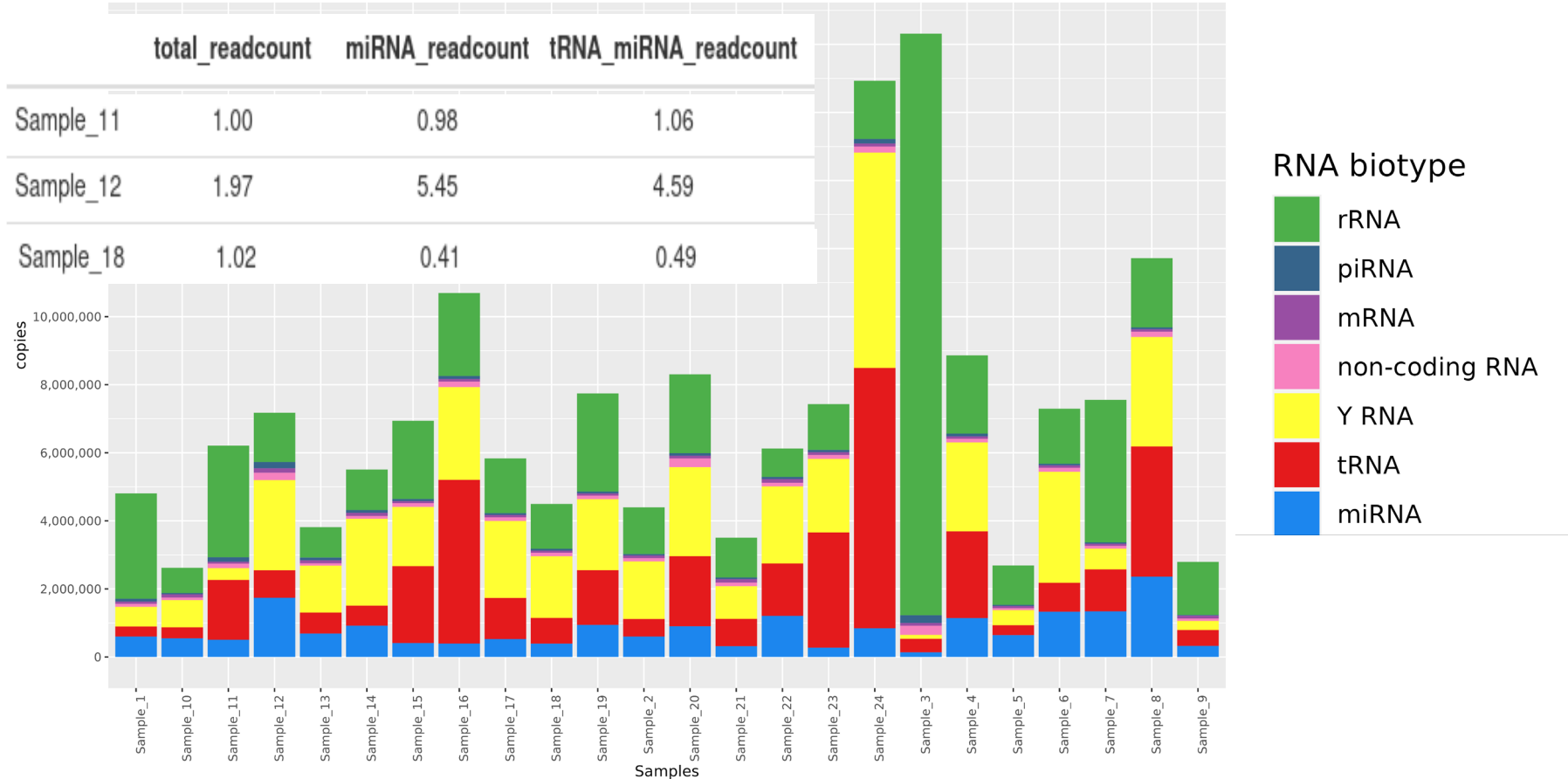
# Library size normalization is skewed by transcript abundance



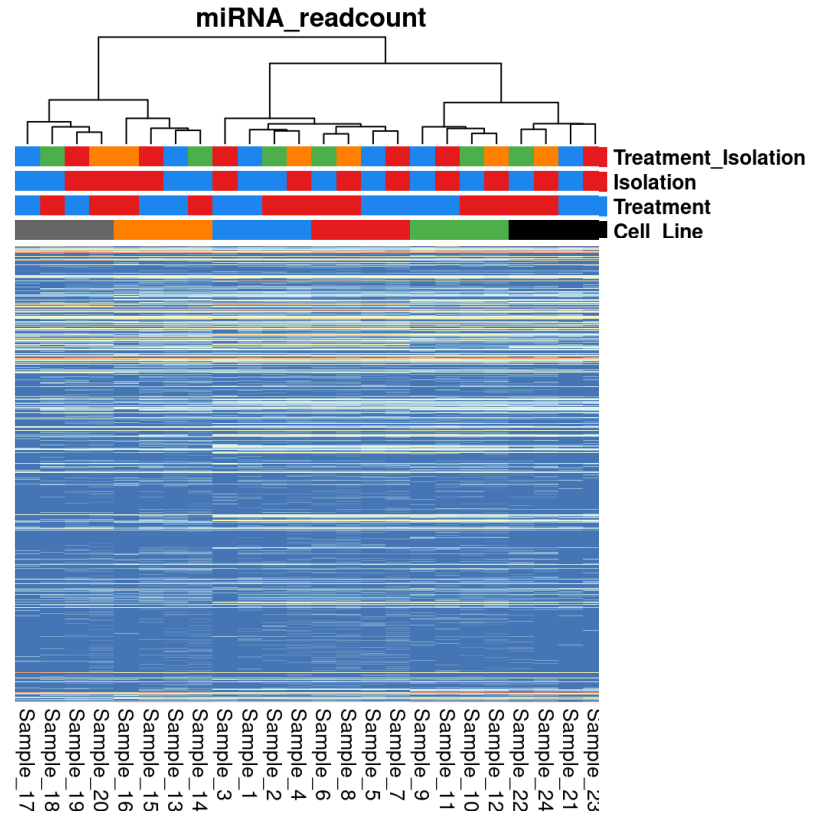
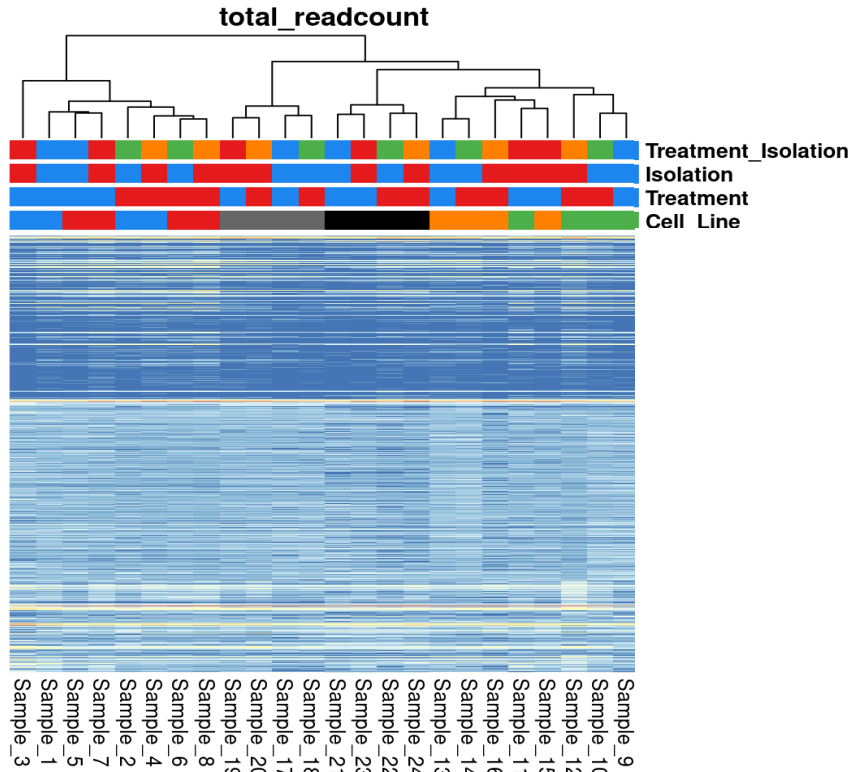
# Median gene expression normalization is skewed by observations



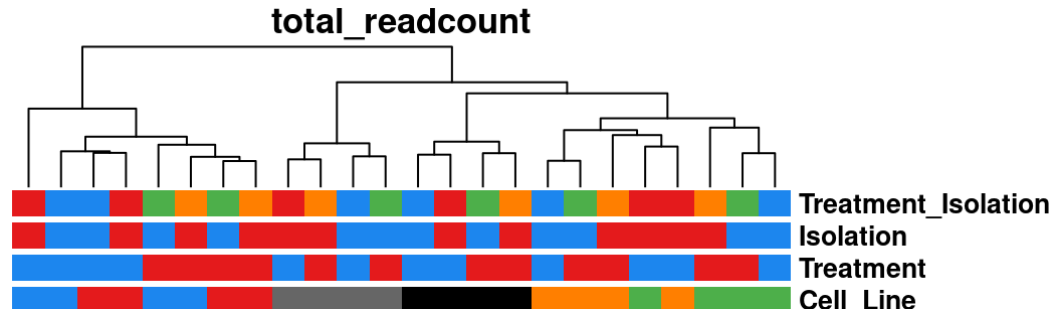
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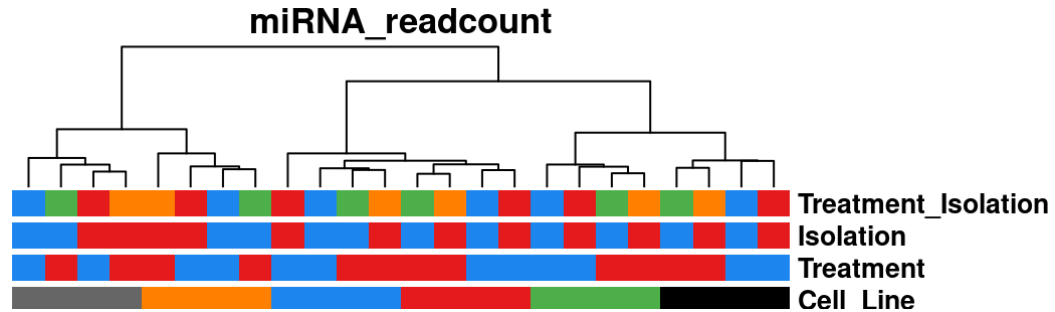
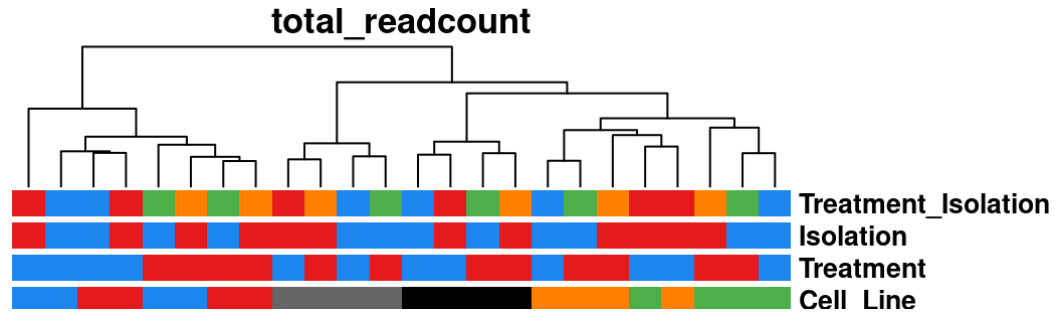
# Normalization can be a major source of bias



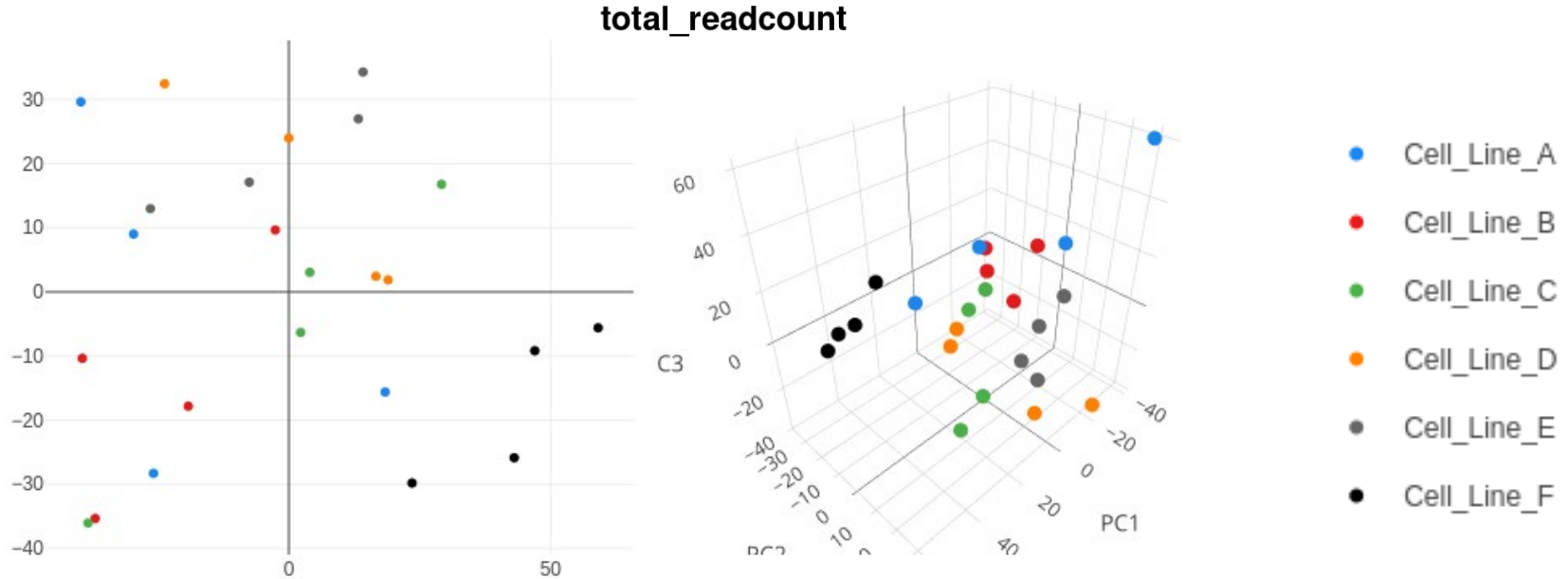
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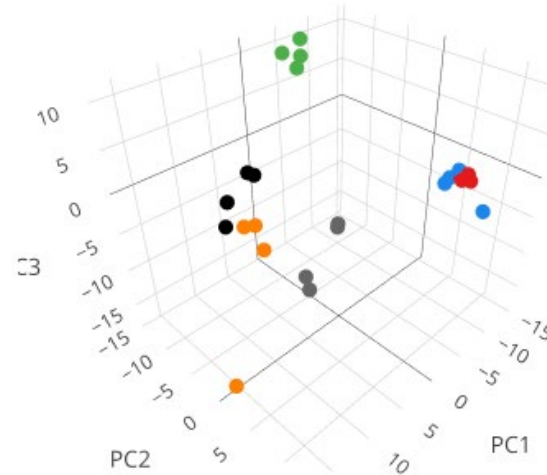
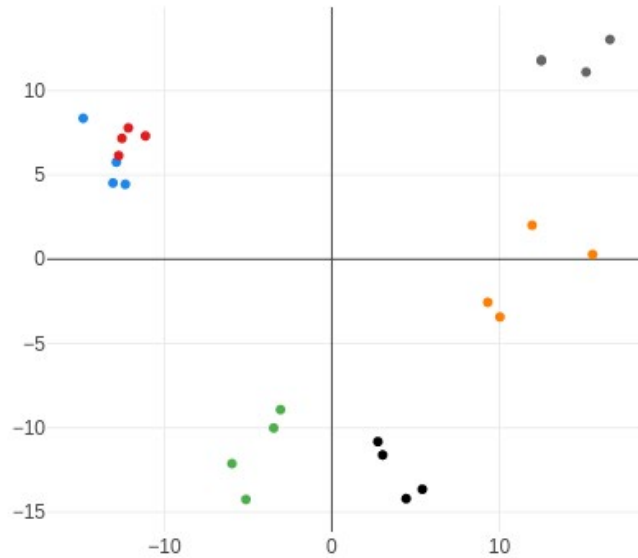


# Correct normalization enables batch effect detection



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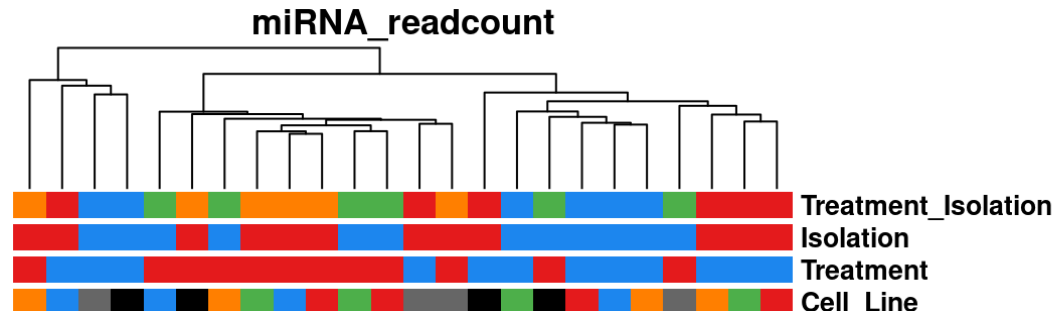
miRNA\_readcount



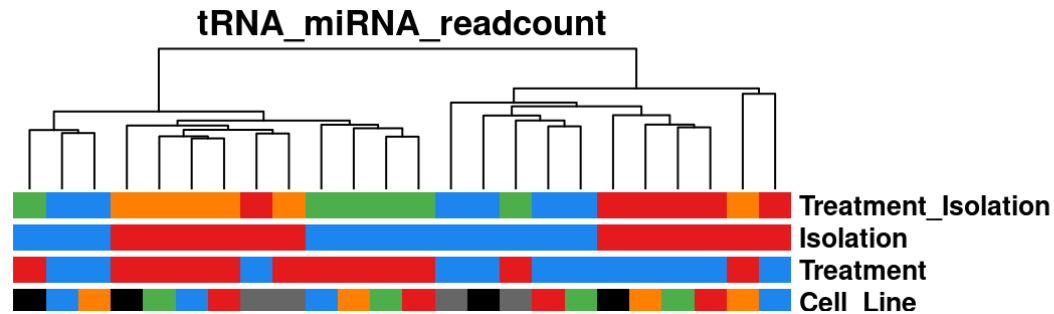
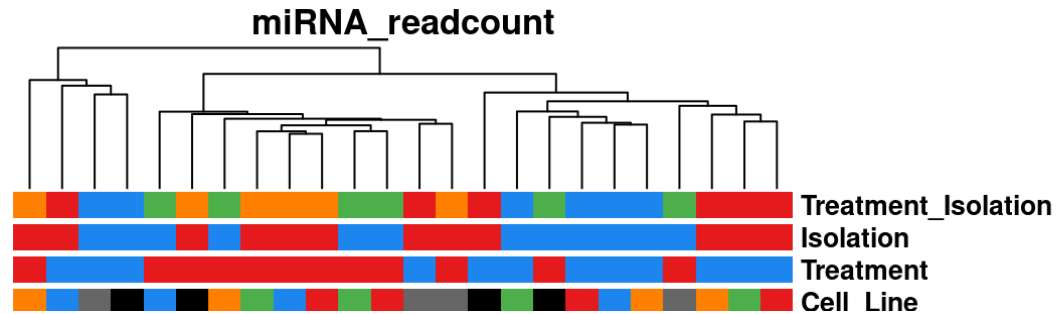
- Cell\_Line\_A
- Cell\_Line\_B
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- Cell\_Line\_D
- Cell\_Line\_E
- Cell\_Line\_F



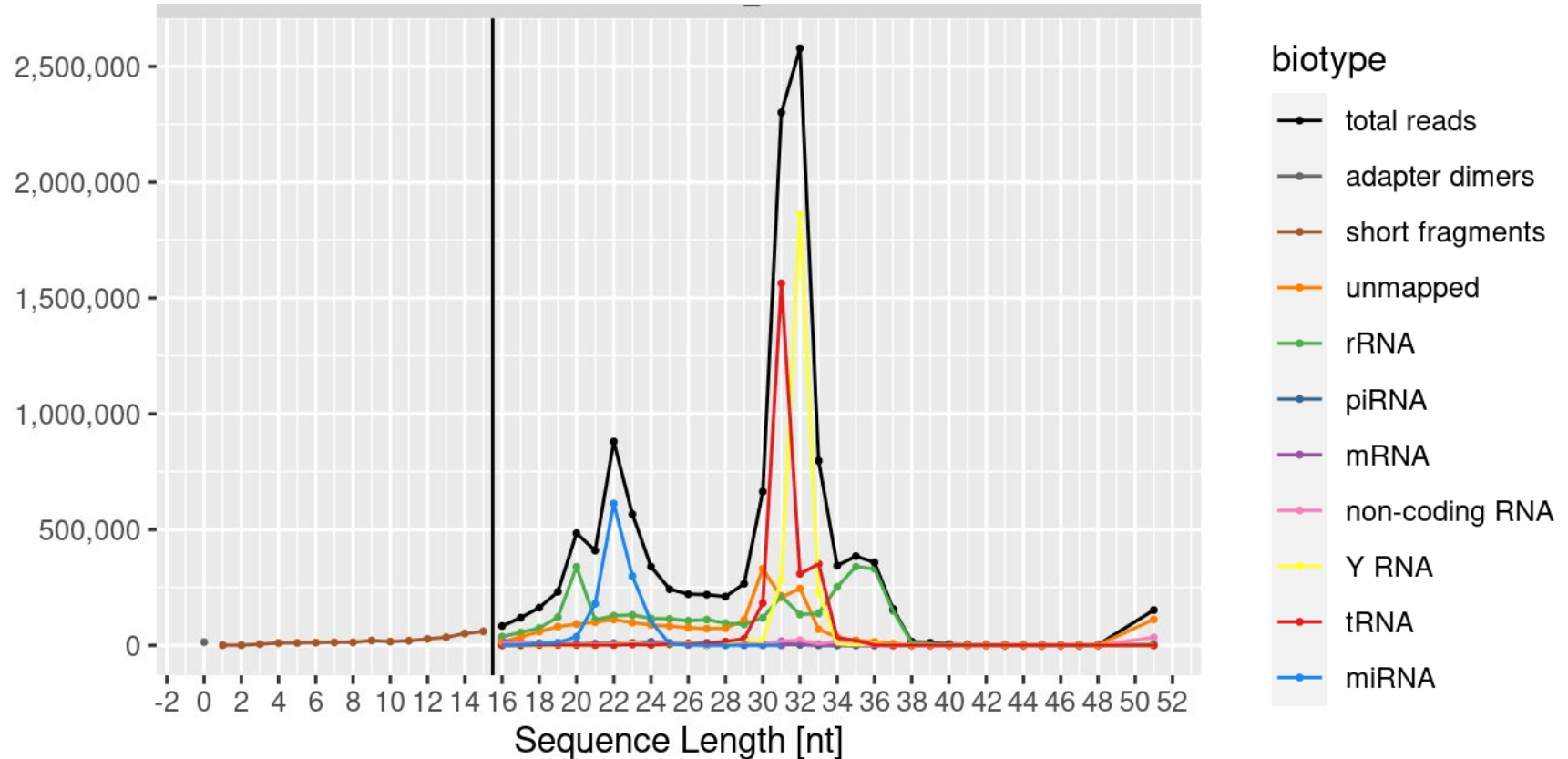
# Exploit small RNA NGS beyond miRNAs



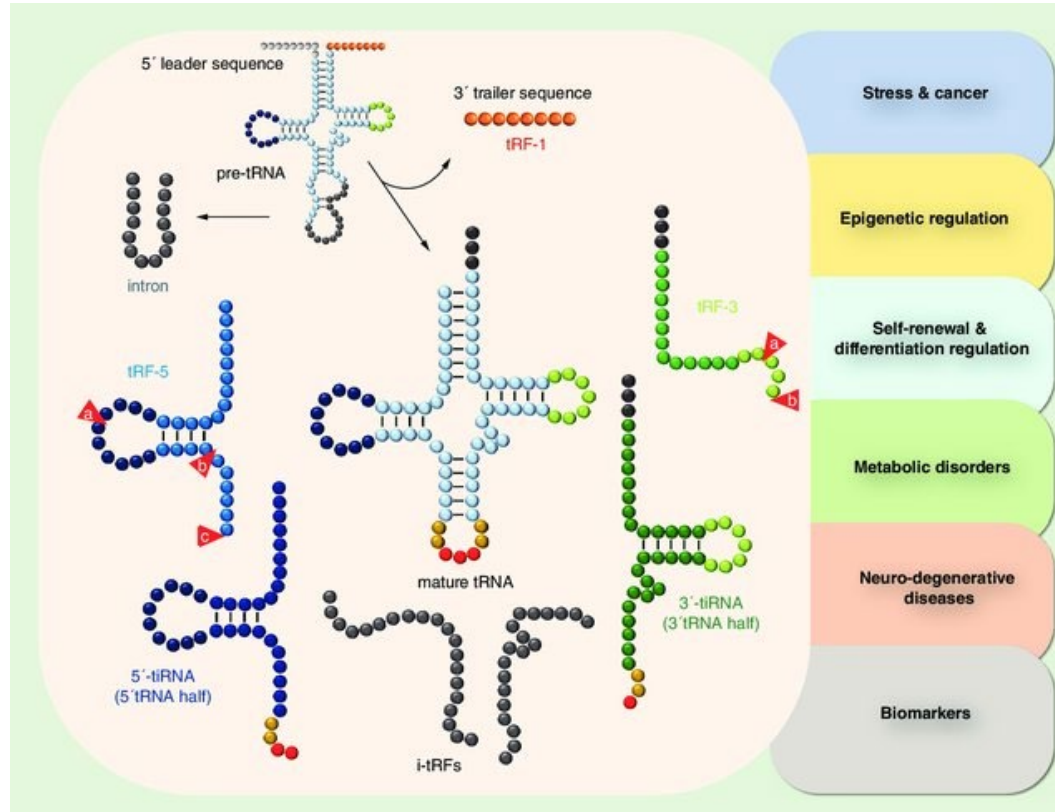
# Improvement of normalization in small RNA NGS



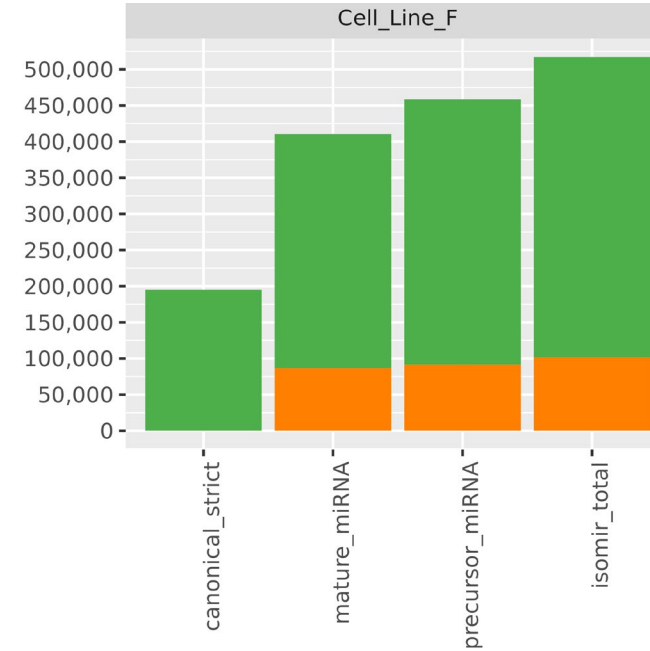
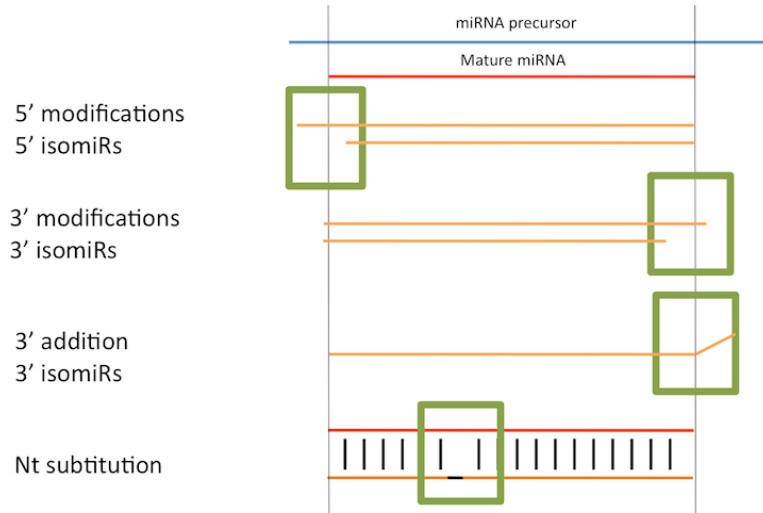
# Degradation vs Specificity in small RNAs



# Hidden complexity in small RNA NGS - tRFs



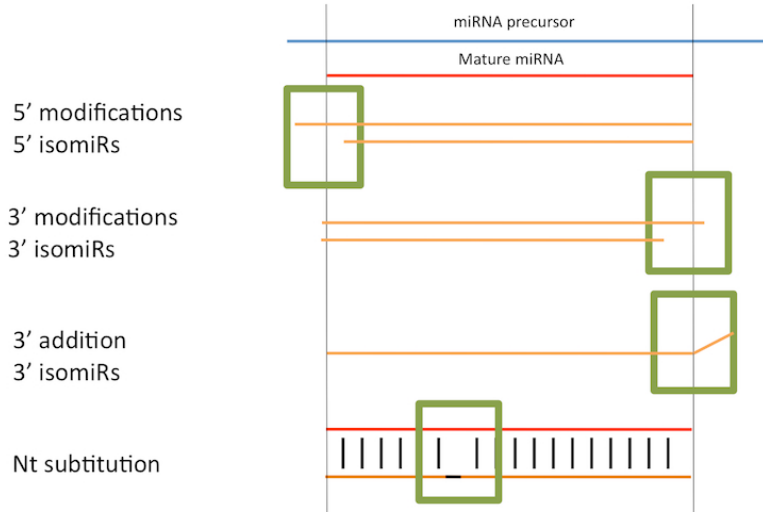
# Hidden complexity in small RNA NGS - isomiRs



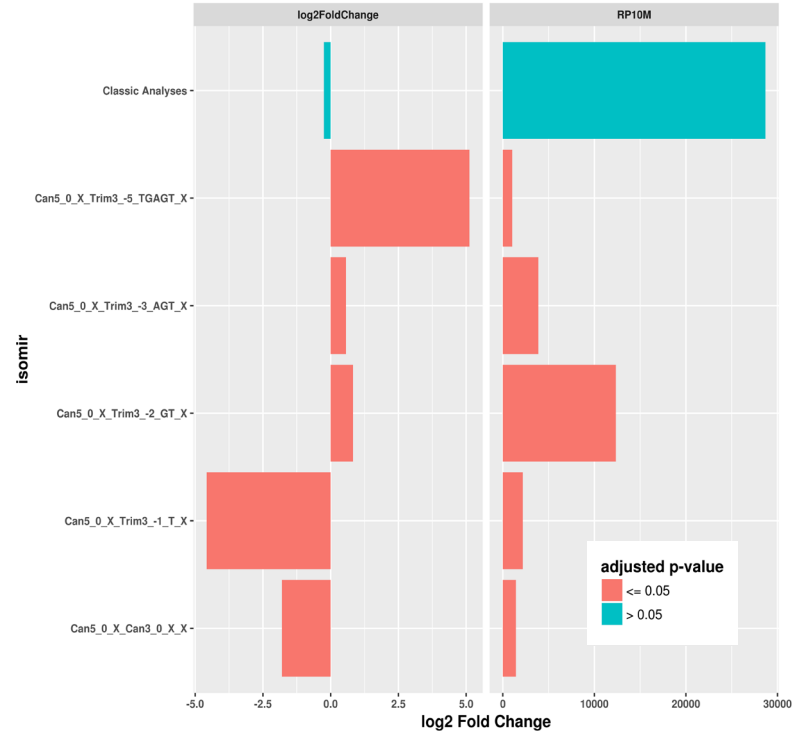
Polymorphic substitutions



# Hidden complexity in small RNA NGS - isomiRs



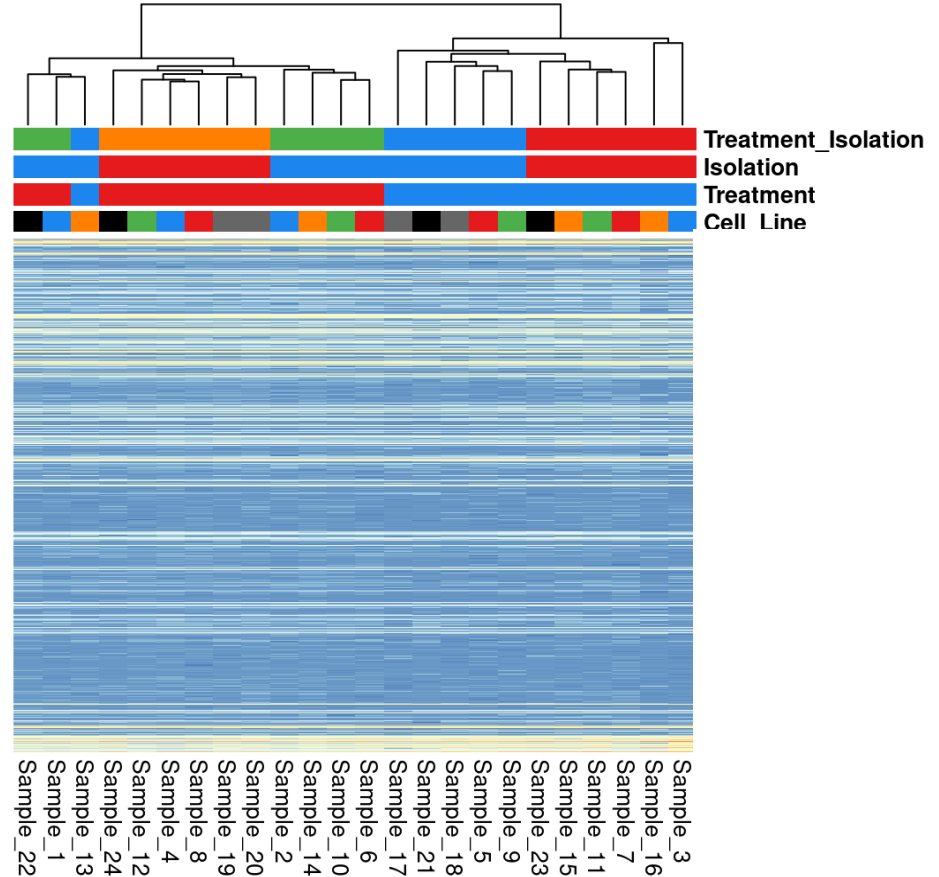
Isomir vs Classic Analyses  
hsa-miR-181a-5p



# Comprehensive analysis of small RNA transcriptome increases efficiency of biomarker detection

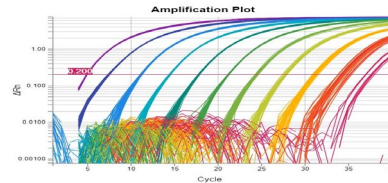


tRFs + isomiRs



# Conclusions

- Importance of spike-ins in small RNA NGS
- Comprehensive characterization of full transcriptome necessary for correct normalization and batch effect detection
- Detection of functional fragments and isoforms increases specificity in classification and biomarker profiles



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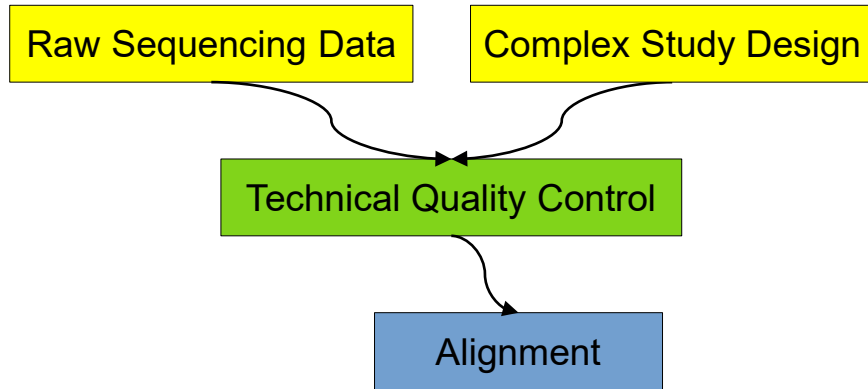


- Detection of stable reference transcripts in small RNA NGS for biomarker validation



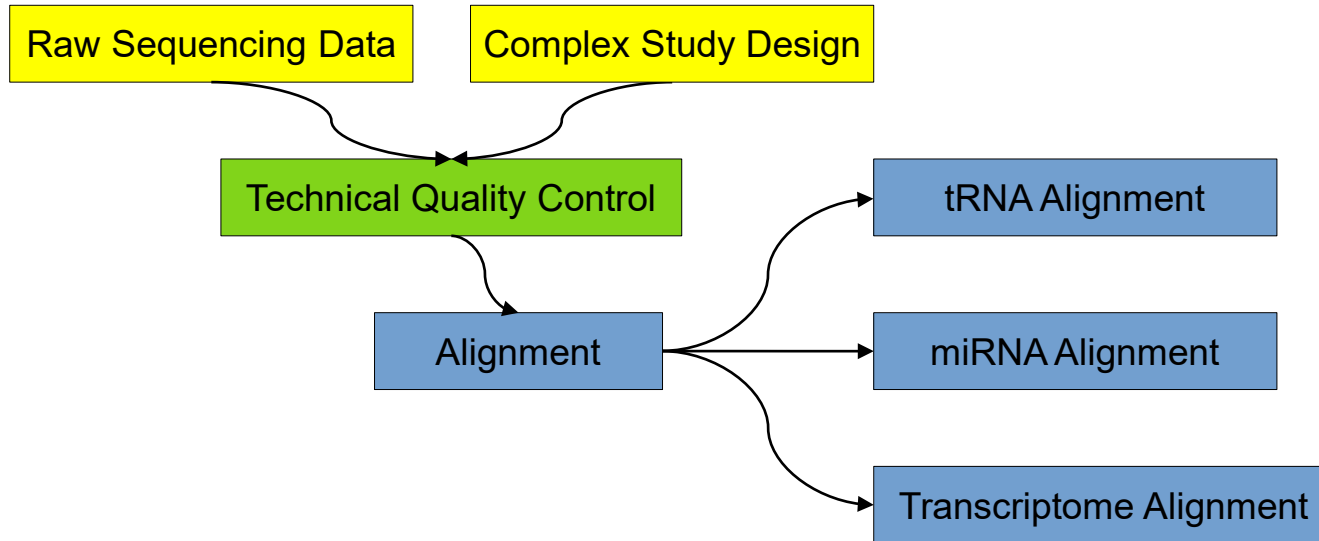
# caRNAge

## Comprehensive Analysis of small RNA Gene Expression



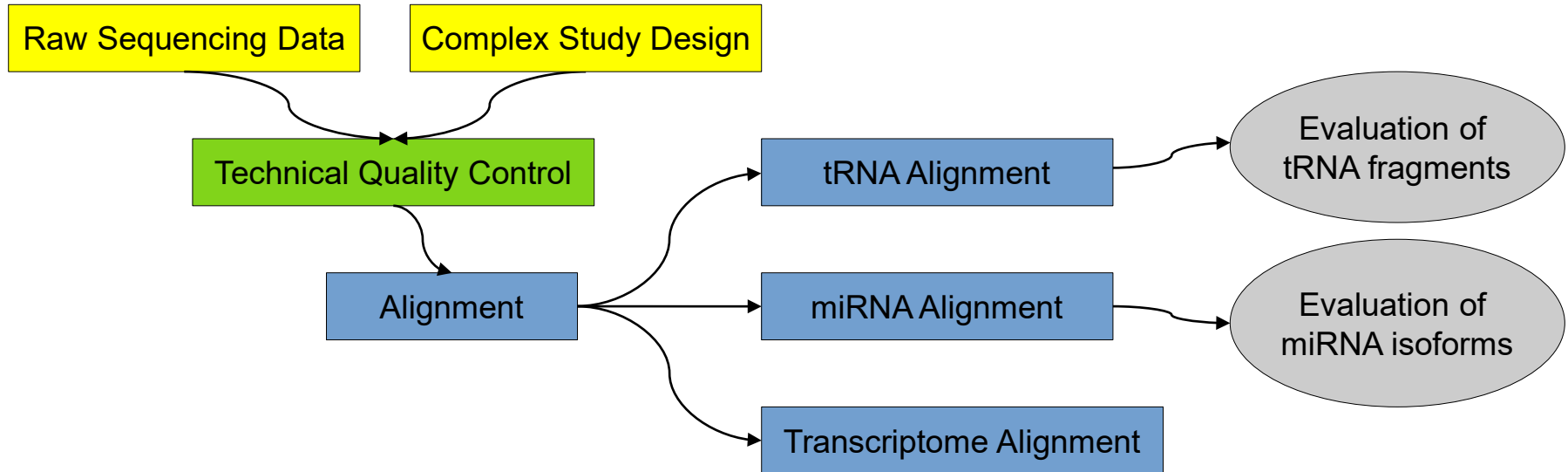
# caRNAge

## Comprehensive Analysis of small RNA Gene Expression



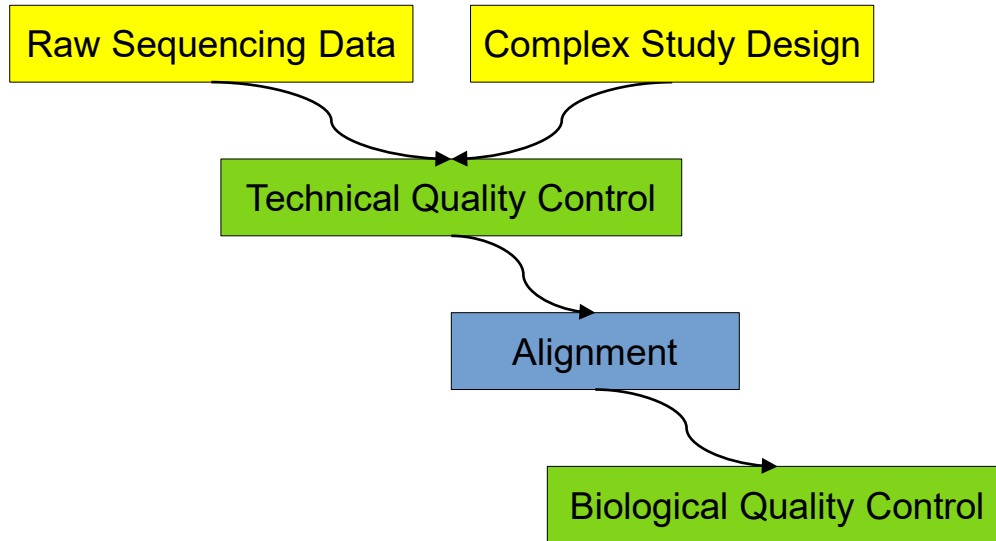
# caRNAge

## Comprehensive Analysis of small RNA Gene Expression



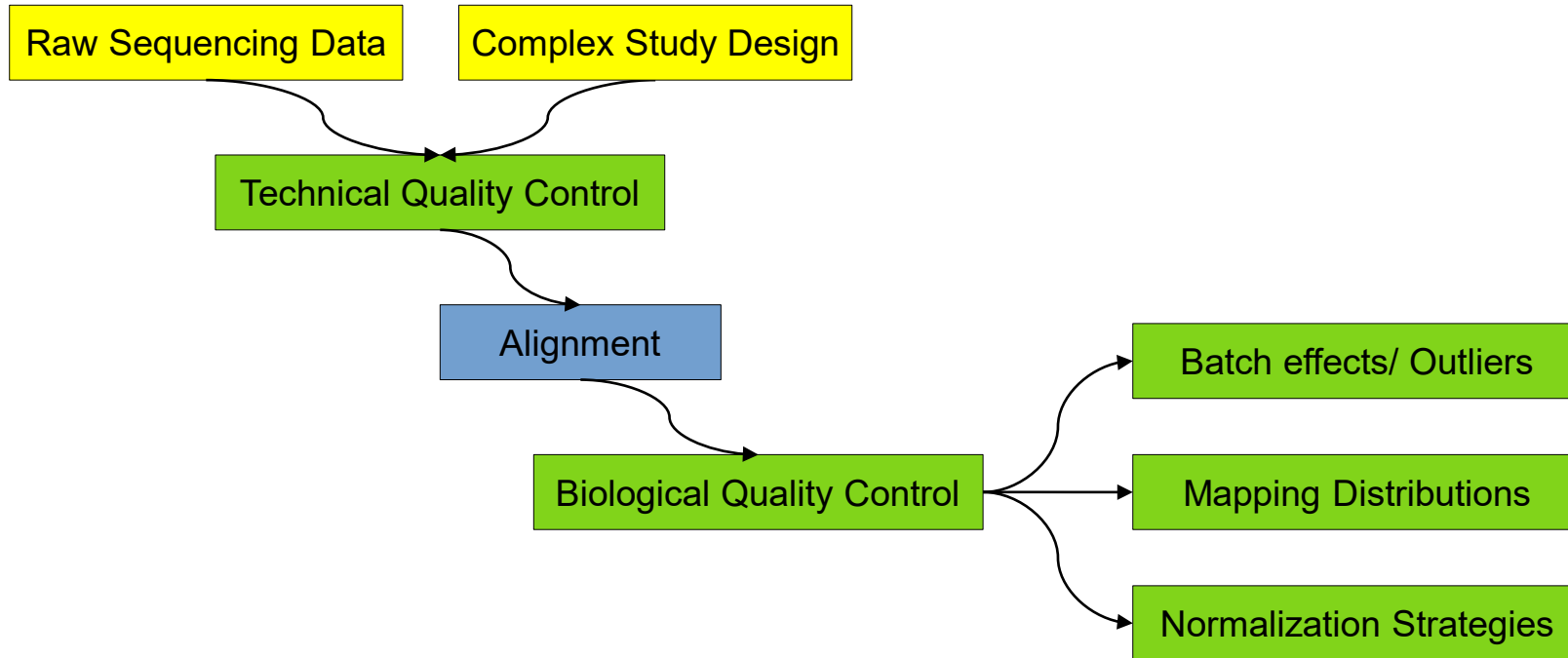
# caRNAge

## Comprehensive Analysis of small RNA Gene Expression



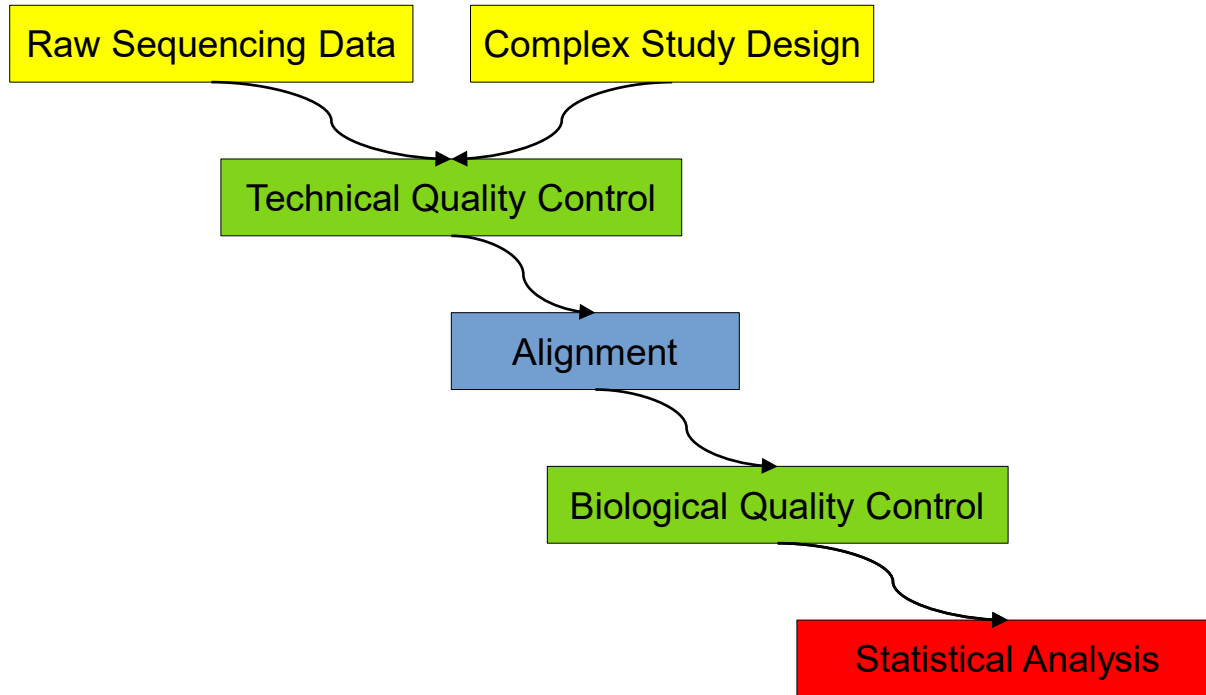
# caRNAge

## Comprehensive Analysis of small RNA Gene Expression



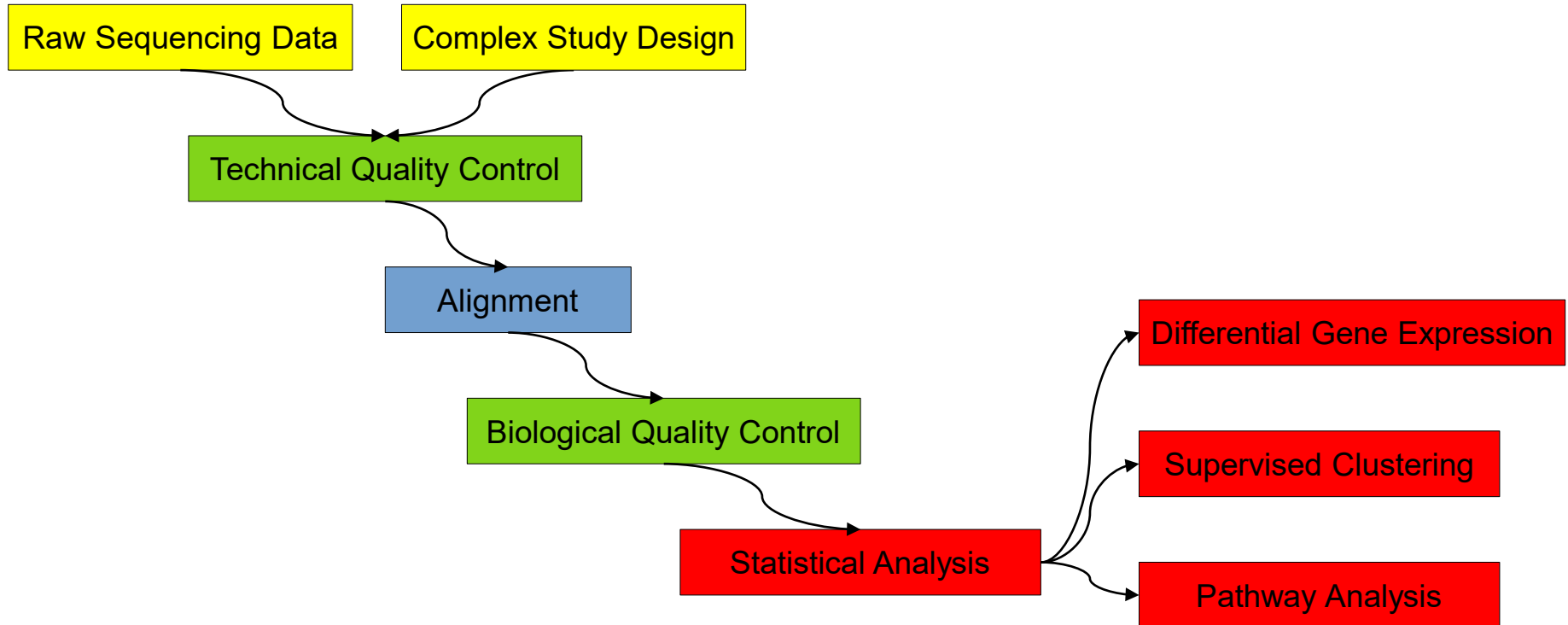
# caRNAge

## Comprehensive Analysis of small RNA Gene Expression



# caRNAge

## Comprehensive Analysis of small RNA Gene Expression



# Output

## Quality Control

- Detection of degradational status
- Detection of sequencing errors
- Detection of batch effects
- Detection of outliers
- Relative distribution of all RNA species
- Length distribution of alignments

## Pathway and gene list analysis

- Identification of predicted and experimentally validated targets of mature miRNAs
- Overrepresentation analysis of significantly regulated transcripts
  - Gene Ontology
  - KEGG
  - Reactome
  - WikiPathways

## Differential gene expression

- Normalization according to specified RNA species
- Statistical evaluation of gene expression changes
- Pairwise comparisons of all specified experimental groups
- Evaluation of stable reference transcripts for qPCR validation

## microRNA and isomiR analysis

- Integrative differential transcript analysis between predominant isomiRs and parent miRNAs
- Analysis of miRNA clusters
- Detection of miRNA localization motifs
- Evaluation of miRNA stability by isomiR modifications

## Supervised clustering

- Detection of optimal error rates
- Sparse projection on latent squares (sPLS)
- Classification based on biomarker profiles



# Acknowledgements

- Division of Animal Physiology and Immunology, TUM
  - Prof. Michael Pfaffl
- Research Group Big Data in Biomedicine, TUM
  - Johannes Kersting
- Institute of Human Genetics, LMU
  - Marlene Reithmair
  - Martina Schuster



Test for yourself!

<https://www.physio.wzw.tum.de/caRNAge/>