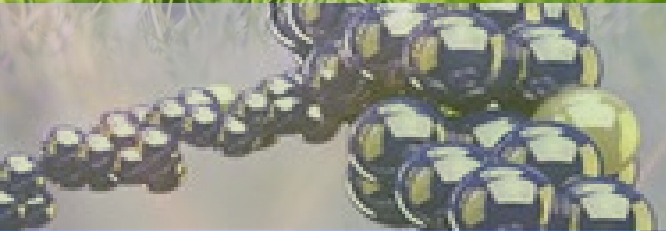




# *Multiomics analysis of pig pre-implantation embryo*

Adrien Dufour, Cyril Kurilo, Jan B. Stöckl, Denis Laloë, Yoann Bailly, Patrick Manceau, Frédéric Martins, Ali G. Turhan, Stéphane Ferchaud, Bertrand Pain, Thomas Fröhlich, Sylvain Foissac, Jérôme Artus, Hervé Acloque



**Micro  
genomics  
2023**

**INRAE**

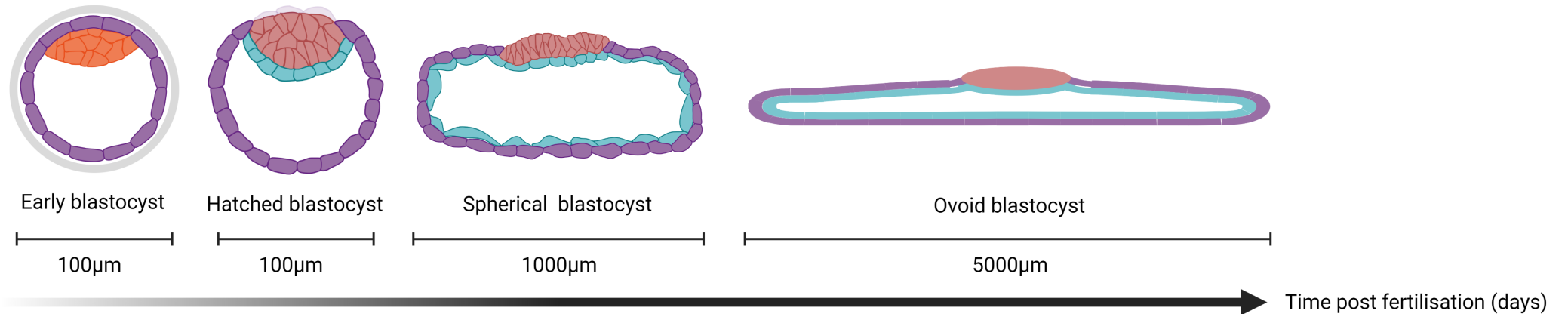


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FIRENZE




# Problematics

- ▶ The use of pluripotent stem cells in livestock is still limited by technological hurdles
- ▶ The optimised modes of obtaining and maintaining pluripotent stem cells for rodents and humans are not adequate for the porcine species

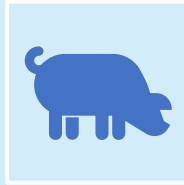
# Working hypothesis



Species	Early blastocyst (100µm)	Hatched blastocyst (100µm)	Spherical blastocyst (1000µm)	Ovoid blastocyst (5000µm)	Implantation (days)
Pig	6	7-8	9-10	11-12	16 = implantation
Cow	6-7	8	9-10	11-13	20 = implantation
Mouse	3.5	4	5.5 = implantation		
Human	5.5	6.5	8 = implantation		

-  Trophectoderm
-  Epiblast (pluripotent cells)
-  Hypoblast

# Working hypothesis

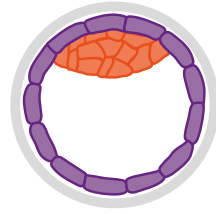


Paracrine regulations specific to pig embryonic development are not taken into consideration for the establishment of a pig's pluripotent lines

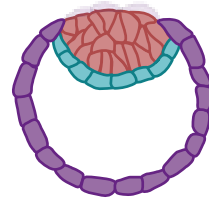


Embryonic pluripotency regulatory networks are not necessarily conserved in mammals

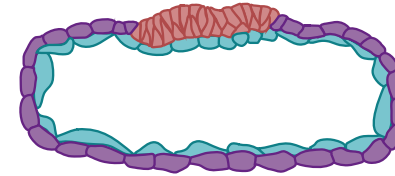
# Methods



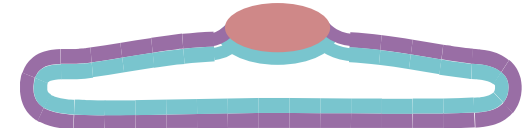
Early blastocyst



Hatched blastocyst



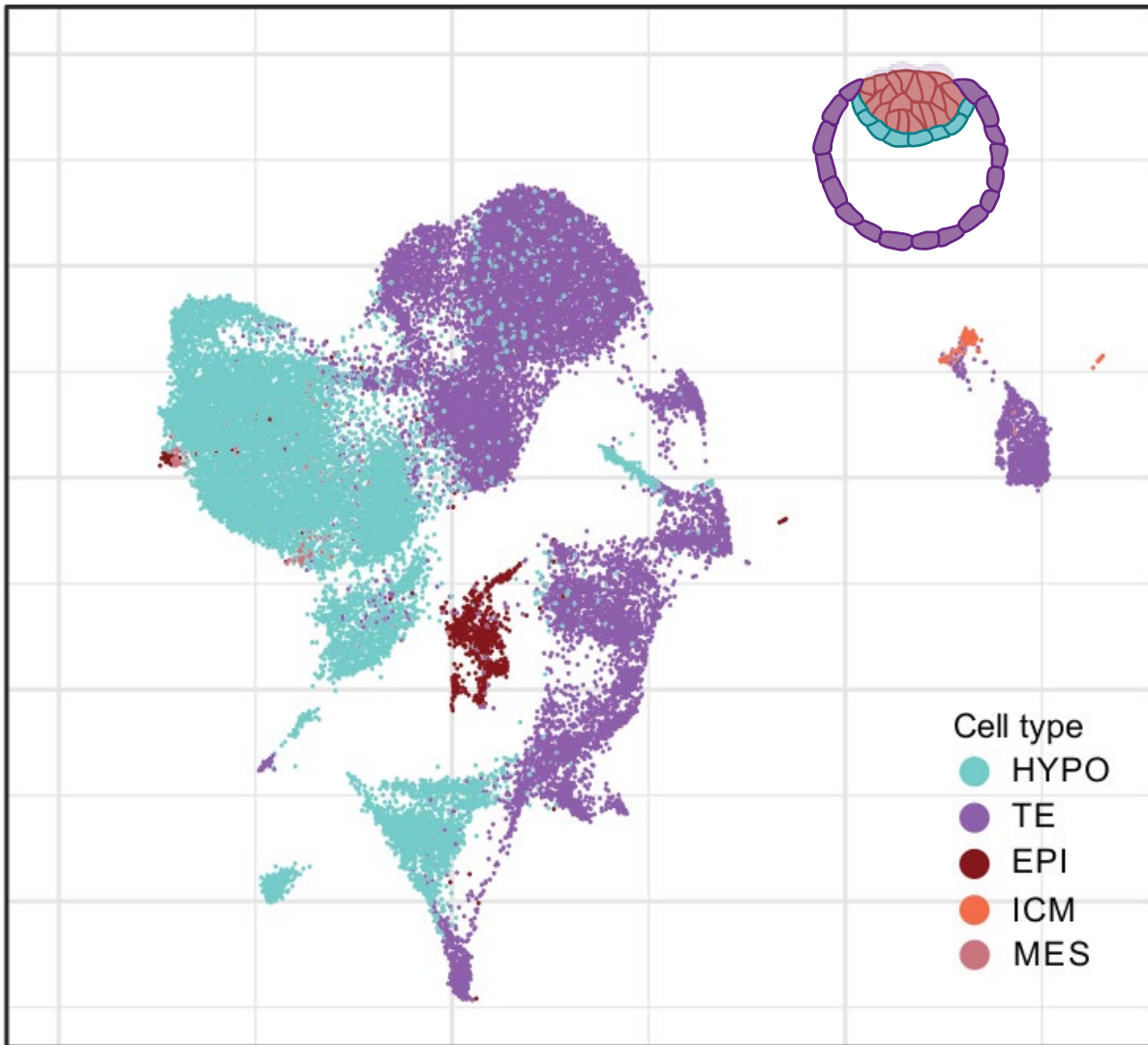
Spherical blastocyst



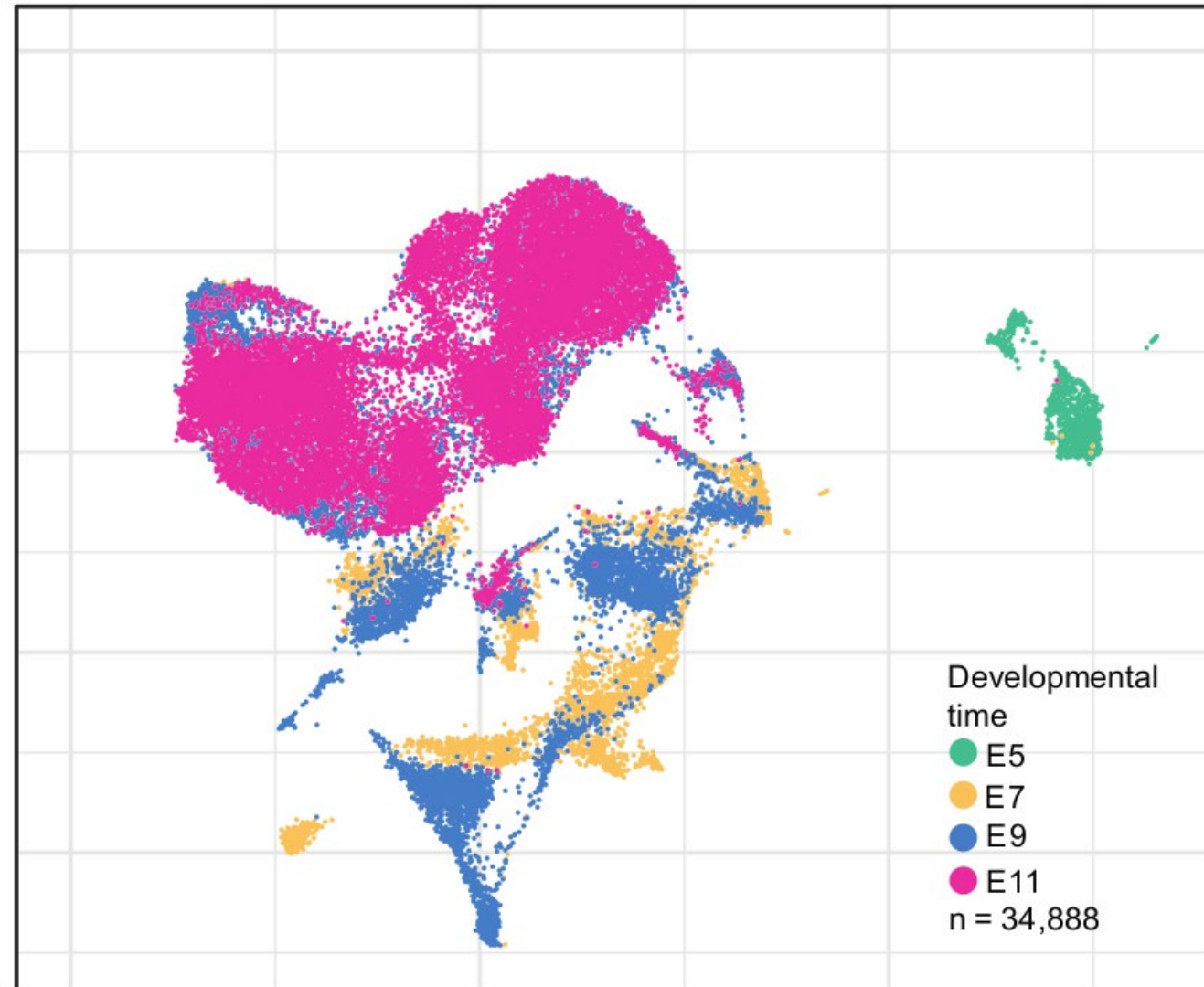
Ovoid blastocyst

Single-cell RNAseq (10x Genomics)	2 libraries (~2000 cells each)	4 libraries (~1000 cells each)	4 libraries (~3000 cells each)	2 libraries (~6000 cells each)
Uterine fluids	8 sows	4 sows	3 sows	3 sows
Single-cell multiomics (10x Genomics) (scATAC-seq + scRNA-seq)	0	1 library (~2000 cell's nuclei)	2 libraries (~4000 cell's nuclei)	4 libraries (~8000 cell's nuclei)

# Identification of the main cell populations



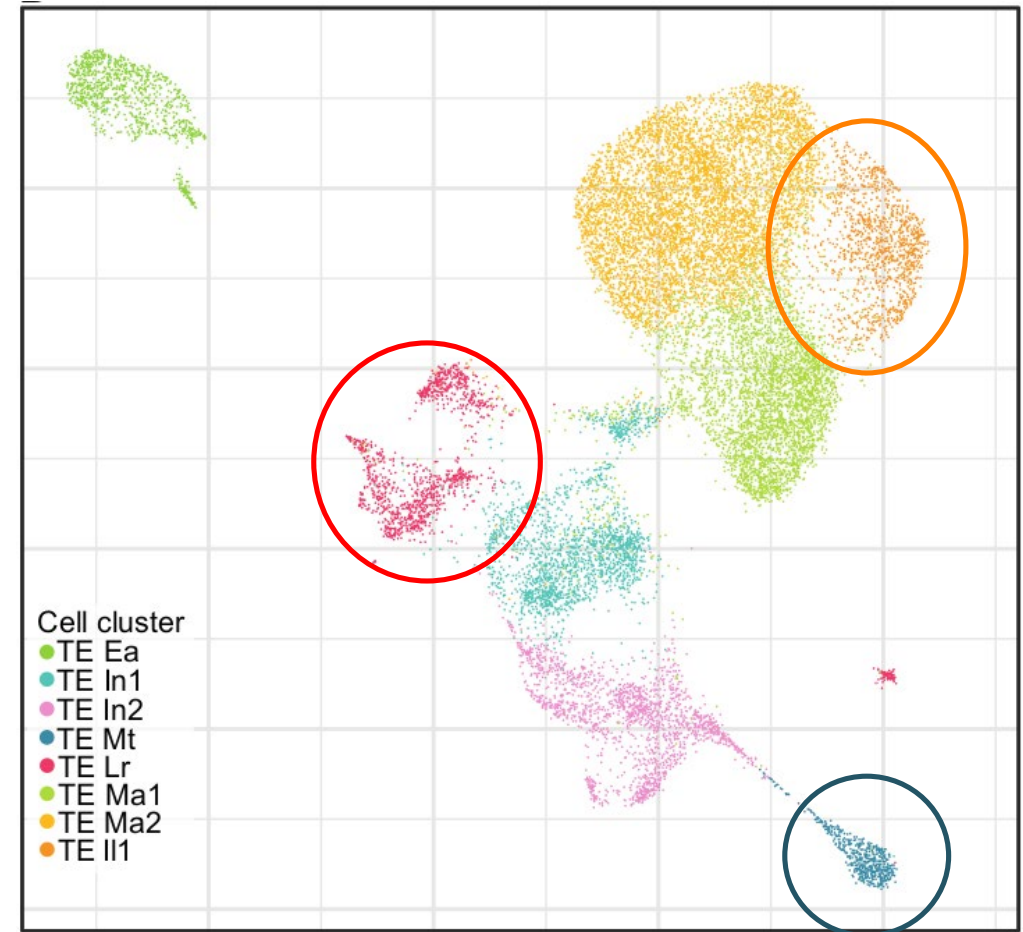
*Visualisation of cells within their lineage*



*Visualisation of cells within their stages*

# Characterisation of sub-populations

- ▶ Our analysis allowed us to characterise sub-populations
- ▶ For the TE, we identified three new subpopulations:
  - ▶ **TE IL1**: IL1 secreting cells
  - ▶ **TE LR**: potential stem cell pop.
  - ▶ **TE Mt**: apoptotic cells that may correspond to the polar TE also named rauber's layer

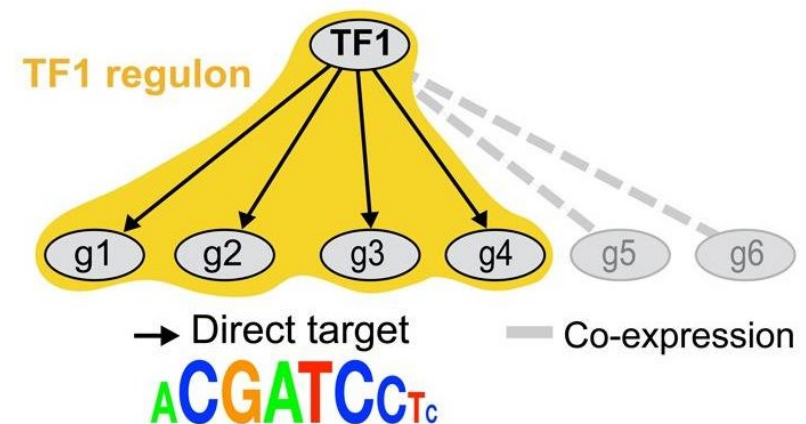


*Visualisation of trophoblast cells within their clusters*

# Identification of modules of regulation (regulon)

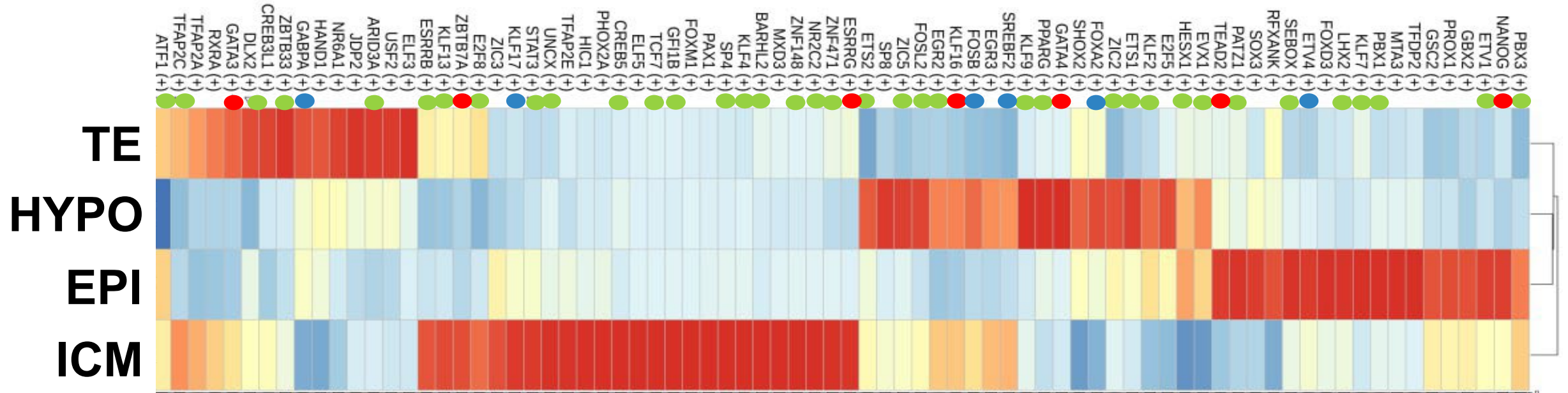
- ▶ Identification of regulons: aggregation of transcription factors and genes with a common TF-binding motif with SCENIC
- ▶ Meta-analyse using human and pig datasets

## Regulons (Gene regulatory network)



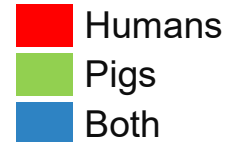


# Identification of modules of regulation (regulon)



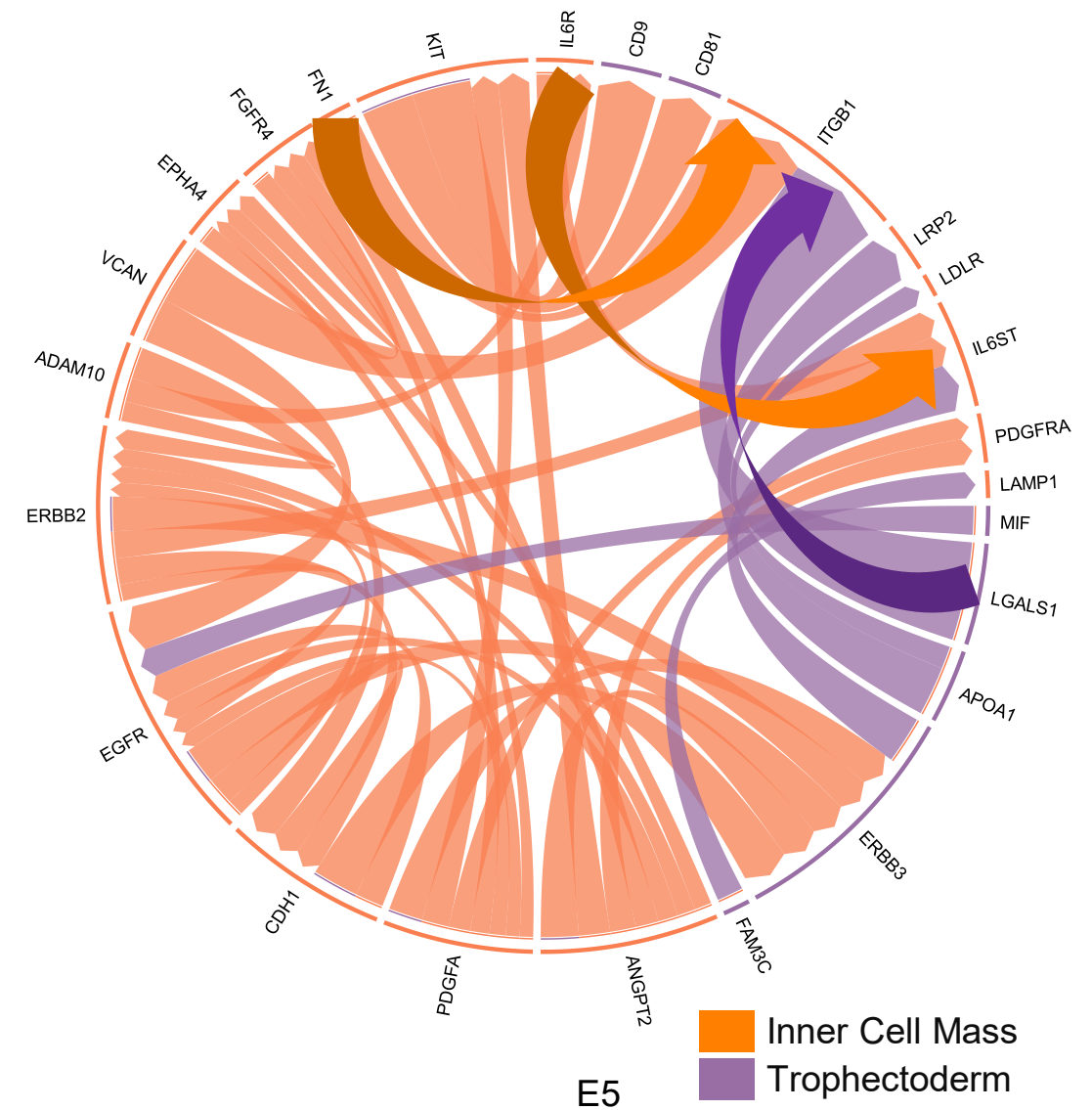
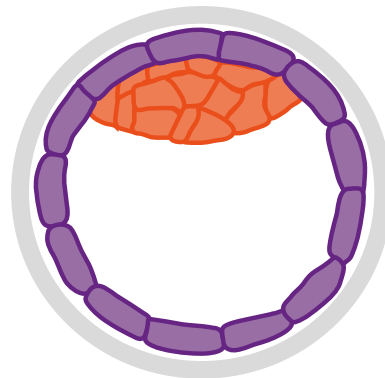
- ▶ New regulons poorly known for the biology of embryonic and extra-embryonic cells: *TFPD2* (EPI), *GABPBA* (TE), *FOSL2* (HYPO)

Commons with :



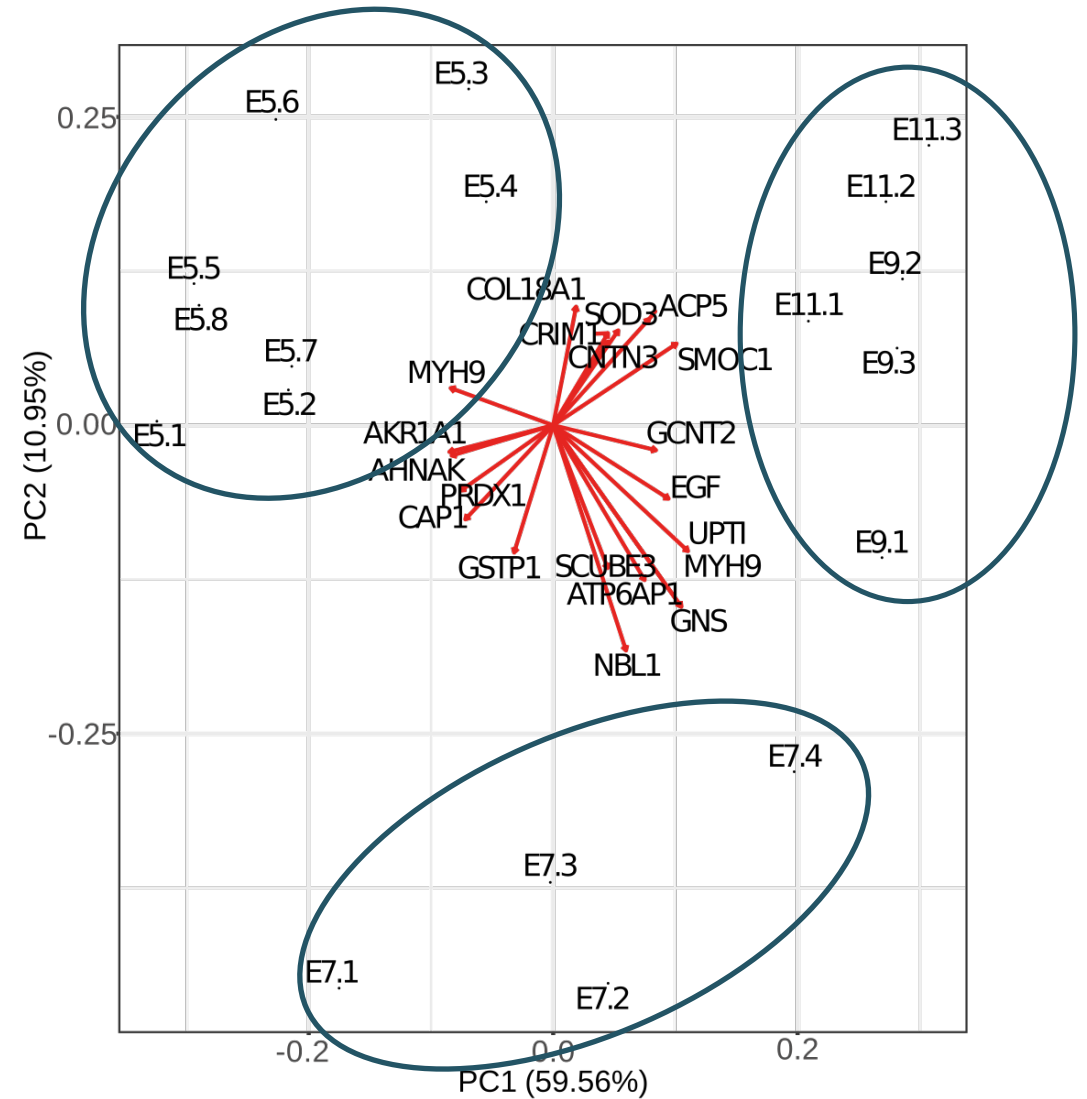
# Identification of Cell-Cell communication

- ▶ Allow us to retrieve known pathways in pigs (Jak-Stat): autocrine
- ▶ Less known pathways like *ITGB1* (autocrine and paracrine)



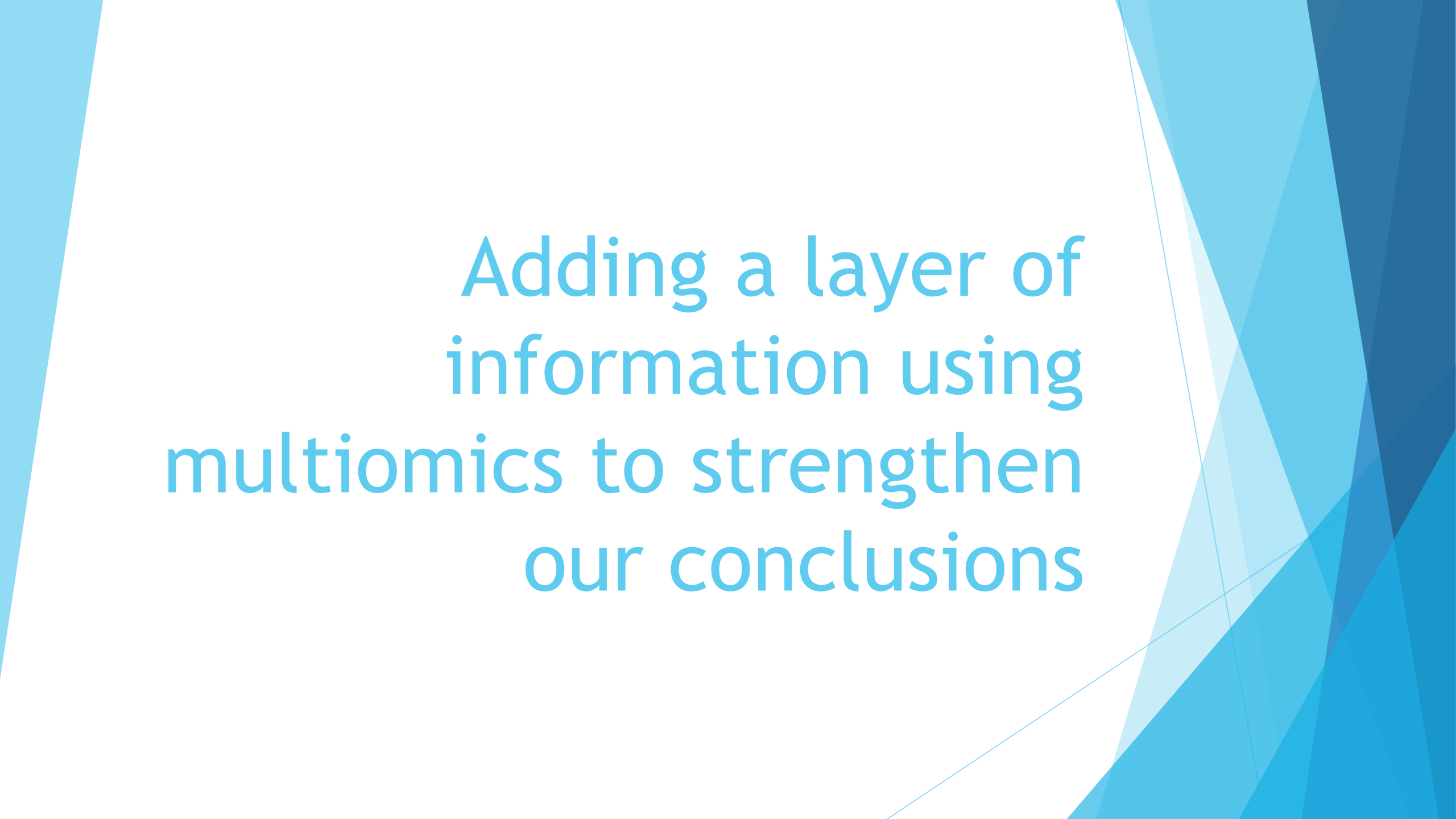
# Proteomics fluids analysis

- ▶ 18 Samples from the same sows as scRNAseq embryos
- ▶ Separation of fluids according to the stage



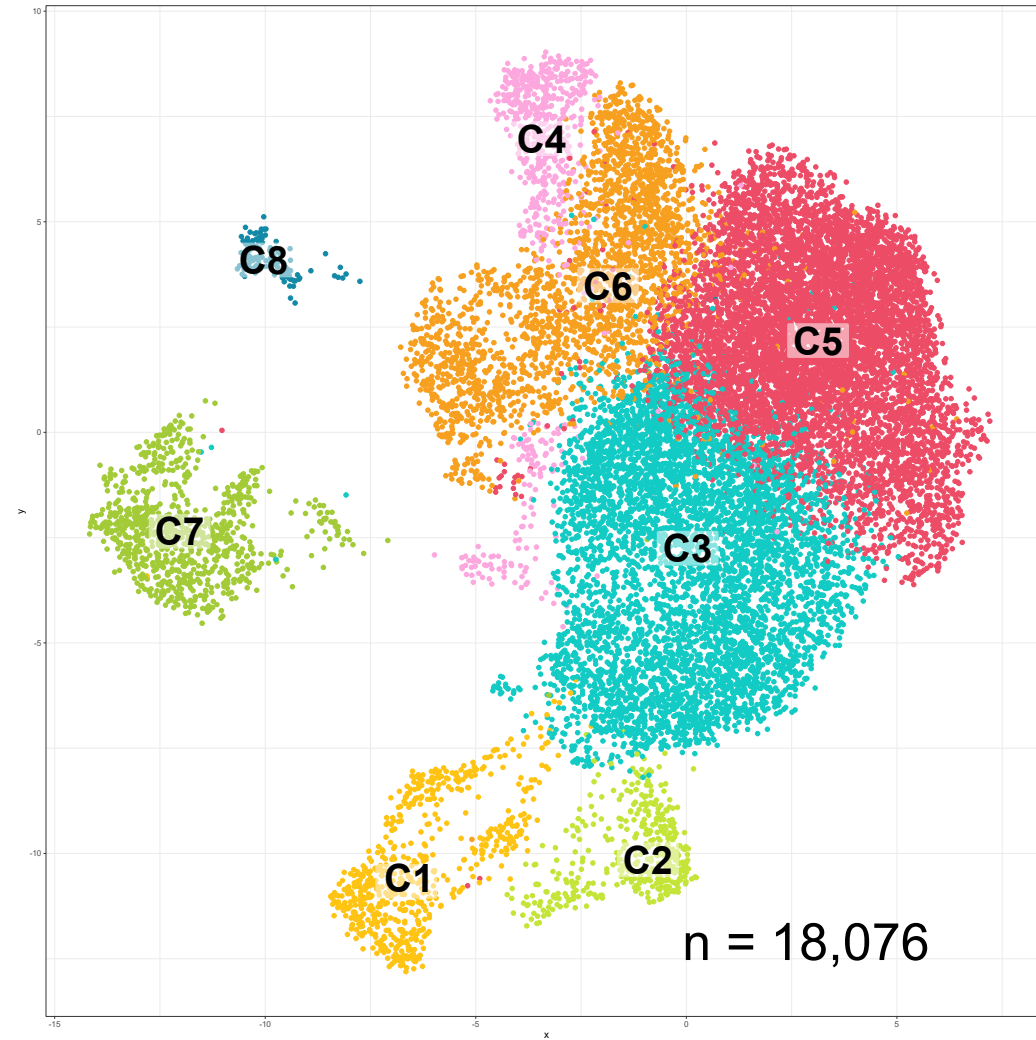
Visualization of uterine fluid samples



The background features abstract, overlapping geometric shapes in various shades of blue, ranging from light sky blue to deep navy blue. The shapes are primarily triangles and polygons, creating a dynamic, layered effect. The text is centered in a clean, sans-serif font.

Adding a layer of  
information using  
multiomics to strengthen  
our conclusions

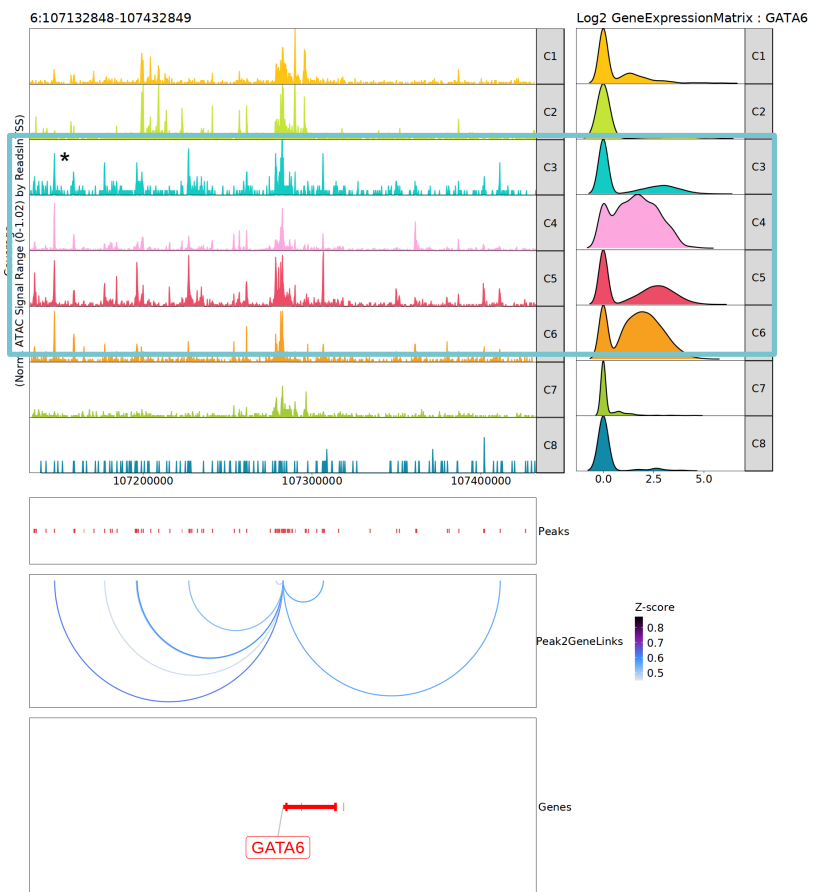
# ScMultiOmics analysis



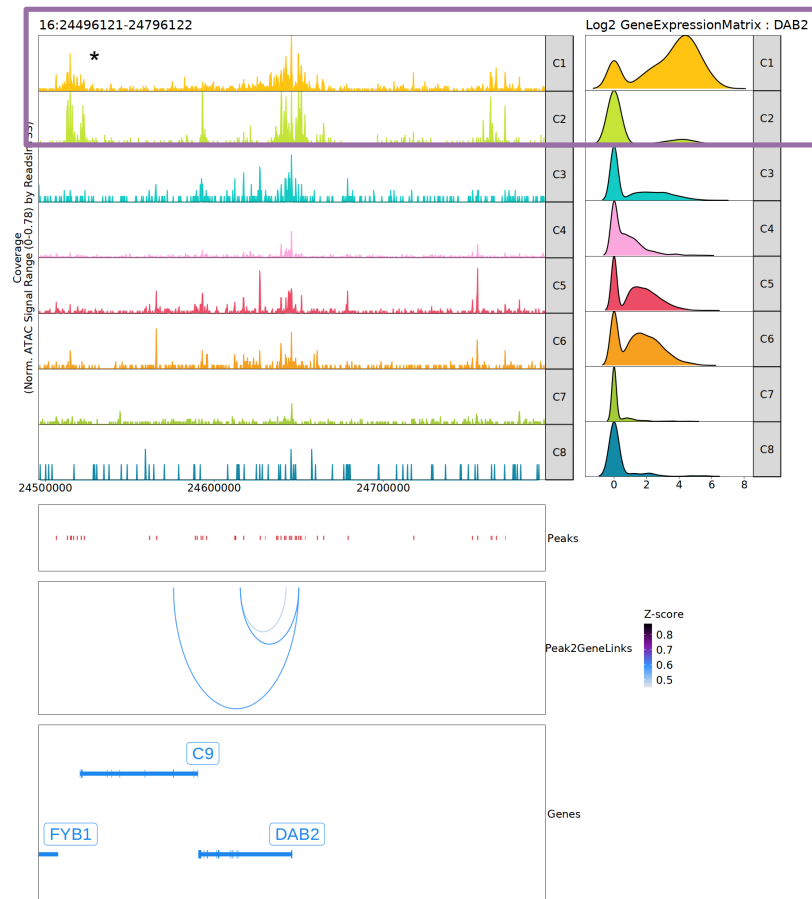
*Visualisation of cells within their clusters*

# Clusters' Identification

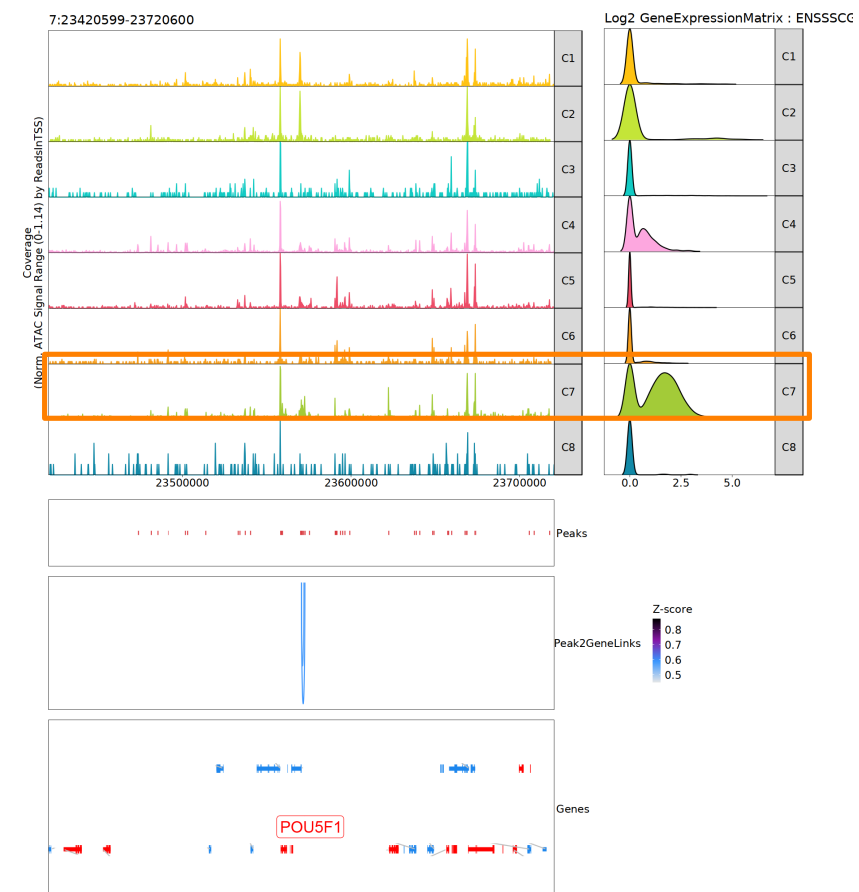
## GATA6 (HYPO)



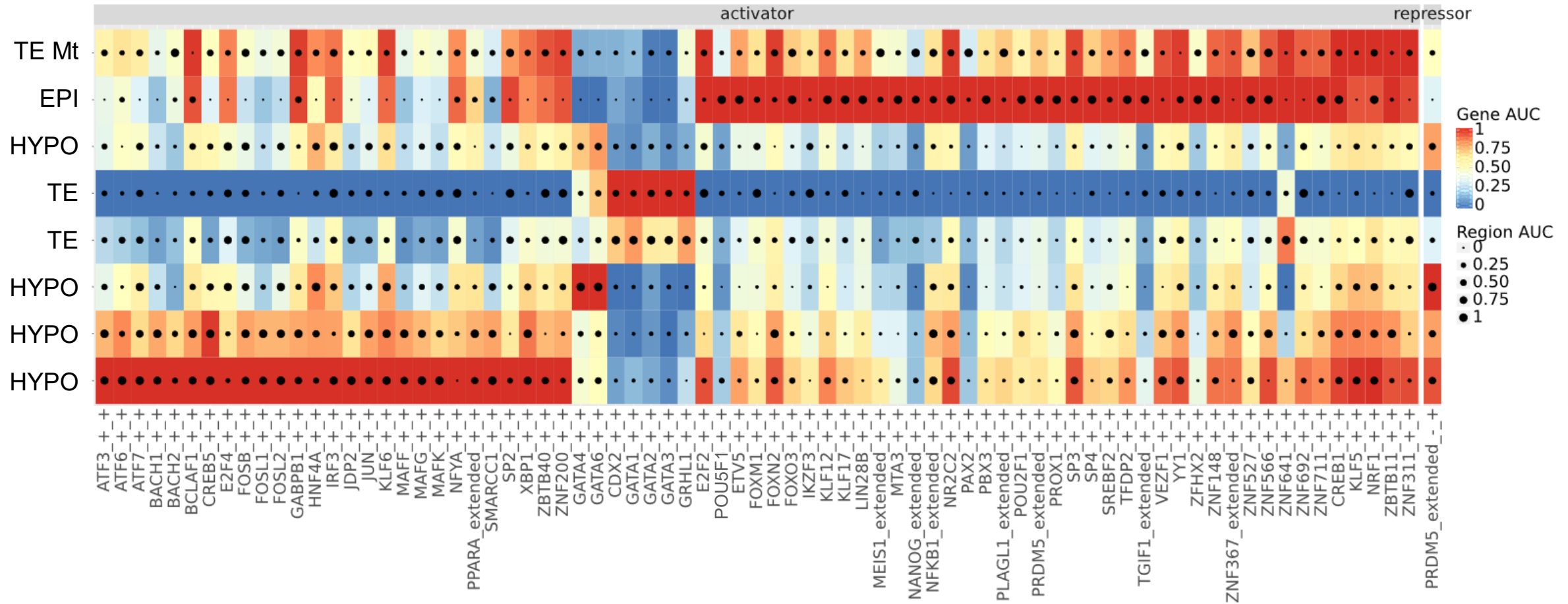
## DAB2 (TE)



## POU5F1 (EPI)



# Identification of regulons using SCENIC+



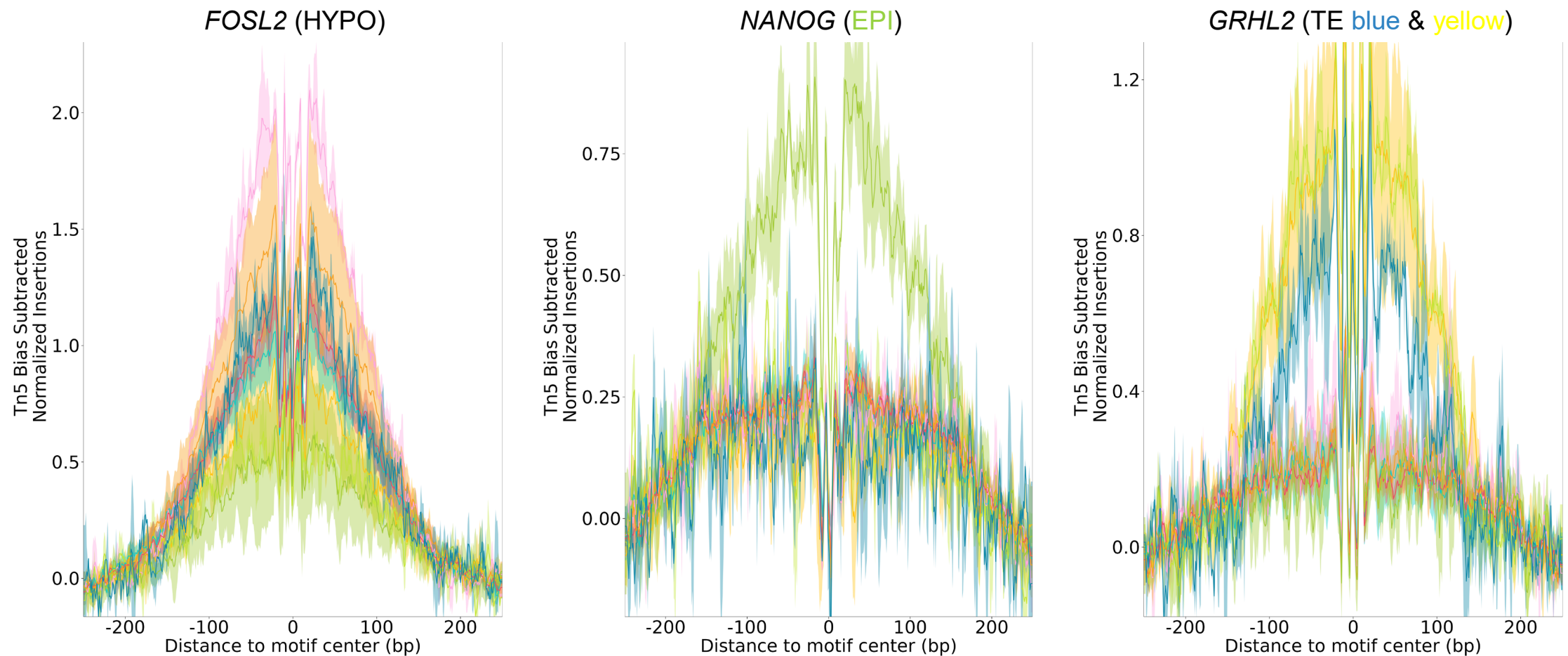
Activity score of regulons identified in terms of decay and gene expression

- ▶ We refined our previously identified (65% similarity) and literature knowledge (*CDX2*, *POU5F1*...)
- ▶ We connect those regulons to regions accessibility



# Footprints confirm the presence of key TFs for each cell population

- ▶ Footprint match eRegulons and literature knowledge
- ▶ Give another level of detail with TF persistence



# Conclusions

- ▶ We identified new TE populations in the pig blastocyst
- ▶ Identification of key paracrine and autocrine interactions between EPI, TE and uterine fluids
- ▶ Multiomics allowed us to refined our regulons identification
- ▶ Better characterisation of regulons with regions accesibility and motif Footprint

# Thanks to

GALAC team and the GABI units

Our collaborators:

- Doriane Guyon
- Cyril Kurylo, GenPHySE UMR1318 INRAE
- Jérôme Artus, UA09 Inserm
- Sylvain Foissac, GenPHySE UMR1318 INRAE
- Yoann Bailly, Patrick Manceau, Stéphane Ferchaud, UE GenESI INRAE
- Thomas Fröhlich, GeneZentrum LMU, Munich, Allemagne

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